

Cell-cell interactions in joint pain: rheumatoid arthritis and osteoarthritis

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Abstract

Rheumatoid and osteoarthritis are chronic conditions generating joint pain for which better management is required. We argue that a better understanding of the cell-to-cell interactions occurring within the joint will enhance mechanistic understanding of joint pain and could lead to new therapeutic avenues being explored.

Text

This Pain Pictured article describes the cell-cell interactions involved in the pathology and pain of rheumatoid arthritis (RA) and osteoarthritis (OA), conditions in which improved pain management is considered key by patients and drives clinical decision making [23,25]. According to the Global Burden of Disease Study 2017, RA had a global prevalence of ~20 million individuals and knee and hip OA ~300 million, both conditions being more common in females than males with prevalence increasing with age, peaking at 60-64 years of age [38,39]. Although both RA and OA cause joint pain with overlap of cellular and inflammatory mediators, there are also key differences in pathology and presentation. For example, RA is an autoimmune disease typically first affecting small joints (i.e. finger/toe joints) but can progress to larger joints, such as the knee, whereas OA pathogenesis involves biomechanical and inflammatory processes affecting multiple joint structures that ultimately lead to structural deterioration with the knee being most commonly afflicted.

Although joint pain is more diffuse than cutaneous pain, most joint structures, including bone, synovium, ligaments, tendon synovial sheaths, infrapatellar fat pad, and menisci are innervated; notably, healthy adult articular cartilage is aneural. Approximately 80% of joint innervating fibers are unmyelinated (a combination of afferent and sympathetic fibers), the remaining 20% being predominantly myelinated A δ -fibers [24,29]. The majority of afferent fibers are nociceptors, a large population being “silent” nociceptors that only become active following sensitization [41].

Joint pathology in RA is characterized by synovial inflammation, cartilage degradation, bone erosion and bone marrow lesions. Synoviocytes proliferate, developing a proinflammatory and catabolic phenotype, whilst numerous immune cells (macrophages, T cells, B cells, plasma cells, mast cells, dendritic cells and neutrophils) infiltrate the synovium and synovial fluid (SF) where they produce proinflammatory mediators, catabolic factors, and chemoattractants that recruit additional immune cells [10,19]. The combined infiltration of T cells, B cells and plasma cells that drive autoimmunity and local autoantibody production is a feature unique to RA when compared to OA. In the OA joint, changes include: alternations in chondrocyte **phenotype**, density and activity – cartilage erosion is followed by chondrocyte hypertrophy and generation of matrix degradation products alongside proinflammatory mediator secretion; infiltration of mast cells, macrophages and CD4+ lymphocytes; synoviocytes proliferate (albeit to a lesser extent than in RA) and release proinflammatory mediators; and changes in subchondral bone turnover due to increased osteoblast and osteoclast activity result in a combination of sclerotic bone, osteophyte formation and subchondral bone marrow lesions with altered vascular and neuronal innervation [5,25,35,46].

Cell-cell interactions occur in RA and OA primarily via release of soluble mediators, the power of which is demonstrated by acellularized human OA-SF sensitizing mouse knee-innervating sensory neurons, thus highlighting the importance of soluble mediators driving cell-cell interactions and pain in arthritis [11]. With regard to the **specific** mediators that drive pain in arthritic joints, there is some overlap between those involved in RA and OA, e.g. non-steroidal anti-inflammatory drugs (NSAIDs) inhibit prostaglandin production and provide relief from pain and inflammation in both conditions. It should be noted however that pain can occur even when inflammation is medically controlled [4]. In both RA and OA, factors that are locally produced in **joints** that could potentially act via nociceptor

expressed receptors include tumor necrosis factor α (TNF α) [26], interleukin 1 β (IL-1 β) [31], IL-6 [16], and IL-17 [15]. While, targeting TNF α , IL-1 β , and IL-6 is efficacious in treating RA, this has not yet been replicated for OA, even though all are implicated in OA pathogenesis. In contrast, nerve growth factor (NGF), which causes hyperalgesia [30], is reportedly elevated in RA and OA SF, and clinical trials highlight anti-NGF antibodies as a promising therapeutic for treating OA pain [7,42], but have yet to be investigated in RA focused clinical trials.

Specifically in OA, toll-like receptor 2 (TLR2), which is activated by a 32-amino-acid aggrecan fragment found in OA-SF, has been highlighted as a potential analgesic target [20,33]. In addition, nociceptors that express voltage-gated sodium channel 1.8 (Na v 1.8) and innervate osteochondral channels have been demonstrated to play a key role in pre-clinical OA models [46], which makes this nociceptor subset a promising therapeutic target considering that nociceptors innervating osteochondral channels are associated with OA pain in humans [5].

Multiple studies indicate that SF pH is decreased in RA, more so than in OA [14,18,21,22], and accordingly, ion channels activated by extracellular protons, such as acid-sensing ion channel 3 (ASIC3) [27,43] and transient receptor potential vanilloid 1 (TRPV1) [9], are linked to mechanical hypersensitivity in experimental arthritis models. **It is however unclear if protons for one specific cell type are responsible for these effects.** Additionally, lipid mediators linked to RA and OA joint inflammation, such as lysophosphatidylcholine, can directly activate ASIC3 [32]. Lastly, in RA, autoantibodies directly stimulate nociceptors by forming immune complexes with cartilage proteins like collagen type II and then activating neuronally expressed Fc γ receptor I [8,36]; autoantibodies can also stimulate other cells to produce pain mediators like CXCL1/2 [45].

In addition to peripheral mechanisms, there are also central changes in RA and OA pain processing [17,28] and **there is substantial debate in the field as to which is clinically more important, but therapeutic targeting of peripheral events is likely to be simpler and associated with fewer side effects.** With regard to central pain processing, in preclinical RA models, expression of TLR4 by spinal microglia and its endogenous ligands, such as HMGB1, has been directly linked to pain-like behavior in a sex and cell specific manner [1,2]. Moreover, TLR4 expressed by local joint immune cells contributes to joint HMGB1-mediated hypersensitivity in males to a larger extent than in females, while TLR4 on nociceptors participates in HMGB1-mediated joint pain in both sexes [37]. Similarly, in preclinical OA models, increased spinal microglia proliferation and activation correlates with pain [40,44]. However, neuroimmune interactions are not restricted to the proximal and distal nerve endings of primary afferent neurons, a key role for macrophage infiltration into the dorsal root ganglia also being illustrated in both OA and RA pain [6,34].

In summary, RA and OA pain involve a multitude of cell-cell interactions and a plethora of proinflammatory mediators, both peripheral and central mechanisms being involved. Increasing mechanistic understanding of the cell-cell interactions occurring and the mediators involved will hopefully lead to identification of new therapeutic strategies, including the potential use of gene therapy to modulate knee-innervating sensory neurons [12].

Conflicts of interest

The authors have no conflicts of interest to declare.

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- [1] Agalave NM, Larsson M, Abdelmoaty S, Su J, Baharpoor A, Lundbäck P, Palmblad K, Andersson U, Harris H, Svensson CI. Spinal HMGB1 induces TLR4-mediated long-lasting

hypersensitivity and glial activation and regulates pain-like behavior in experimental arthritis: *Pain* 2014;155:1802–1813.

- [2] Agalave NM, Rudjito R, Farinotti AB, Khoonsari PE, Sandor K, Nomura Y, Szabo-Pardi TA, Urbina CM, Palada V, Price TJ, Harris HE, Burton MD, Kultima K, Svensson CI. Sex-dependent role of microglia in disulfide HMGB1-mediated mechanical hypersensitivity. *Pain* 2020.
- [3] Aloe L, Tuveri MA, Carcassi U, Levi-Montalcini R. Nerve growth factor in the synovial fluid of patients with chronic arthritis. *Arth Rheum* 1992;35:351–355.
- [4] Altawil R, Saevarsdottir S, Wedrén S, Alfredsson L, Klareskog L, Lampa J. Remaining Pain in Early Rheumatoid Arthritis Patients Treated With Methotrexate: Methotrexate and Pain in Early RA Patients. *Arthritis Care & Research* 2016;68:1061–1068.
- [5] Aso K, Shahtaheri SM, Hill R, Wilson D, McWilliams DF, Nwosu LN, Chapman V, Walsh DA. Contribution of nerves within osteochondral channels to osteoarthritis knee pain in humans and rats. *Osteoarthritis and Cartilage* 2020:S1063458420310268.
- [6] von Banchet G, Boettger MK, Fischer N, Gajda M, Bräuer R, Schaible H-G. Experimental arthritis causes tumor necrosis factor- α -dependent infiltration of macrophages into rat dorsal root ganglia which correlates with pain-related behavior: *Pain* 2009;145:151–159.
- [7] Berenbaum F, Blanco FJ, Guermazi A, Miki K, Yamabe T, Viktrup L, Junor R, Carey W, Brown MT, West CR, Verburg KM. Subcutaneous tanezumab for osteoarthritis of the hip or knee: efficacy and safety results from a 24-week randomised phase III study with a 24-week follow-up period. *Ann Rheum Dis* 2020;79:800–810.
- [8] Bersellini Farinotti A, Wigerblad G, Nascimento D, Bas DB, Morado Urbina C, Nandakumar KS, Sandor K, Xu B, Abdelmoaty S, Hunt MA, Ängeby Möller K, Baharpoor A, Sinclair J, Jardemark K, Lanner JT, Khmaladze I, Borm LE, Zhang L, Wermeling F, Cragg MS, Lengqvist J, Chabot-Doré A-J, Diatchenko L, Belfer I, Collin M, Kultima K, Heyman B, Jimenez-Andrade JM, Codeluppi S, Holmdahl R, Svensson CI. Cartilage-binding antibodies induce pain through immune complex-mediated activation of neurons. *Journal of Experimental Medicine* 2019;216:1904–1924.
- [9] Borbély É, Botz B, Bölcskei K, Kenyér T, Kereskai L, Kiss T, Szolcsányi J, Pintér E, Csepregi JZ, Mócsai A, Helyes Z. Capsaicin-sensitive sensory nerves exert complex regulatory functions in the serum-transfer mouse model of autoimmune arthritis. *Brain, Behavior, and Immunity* 2015;45:50–59.
- [10] Catrina AI, Svensson CI, Malmström V, Schett G, Klareskog L. Mechanisms leading from systemic autoimmunity to joint-specific disease in rheumatoid arthritis. *Nat Rev Rheumatol* 2017;13:79–86.
- [11] Chakrabarti S, Jadon DR, Bulmer DC, Smith ESJ. Human osteoarthritic synovial fluid increases excitability of mouse dorsal root ganglion sensory neurons: an in-vitro translational model to study arthritic pain. *Rheumatol* 2020;59:662–667.
- [12] Chakrabarti S, Pattison LA, Doleschall B, Rickman RH, Blake H, Callejo G, Heppenstall PA, Smith ESJ. Intra-articular AAV-PHP.S mediated chemogenetic targeting of knee-innervating dorsal root ganglion neurons alleviates inflammatory pain in mice. *Arthritis & Rheumatology (Hoboken, NJ)* 2020.
- [13] Craig AD, Heppelmann B, Schaible H-G. The projection of the medial and posterior articular nerves of the cat's knee to the spinal cord. *J Comp Neurol* 1988;276:279–288.

- [14] Cummings NA, Nordby GL. Measurement of synovial fluid pH in normal and arthritic knees. *Arthritis & Rheumatism* 1966;9:47–56.
- [15] Ebbinghaus M, Natura G, Segond von Banchet G, Hensellek S, Böttcher M, Hoffmann B, Salah FS, Gajda M, Kamradt T, Schaible H-G. Interleukin-17A is involved in mechanical hyperalgesia but not in the severity of murine antigen-induced arthritis. *Sci Rep* 2017;7:10334.
- [16] Ebbinghaus M, Segond von Banchet G, Massier J, Gajda M, Bräuer R, Kress M, Schaible H-G. Interleukin-6-dependent influence of nociceptive sensory neurons on antigen-induced arthritis. *Arthritis Res Ther* 2015;17:334.
- [17] Eitner A, Hofmann GO, Schaible H-G. Mechanisms of Osteoarthritic Pain. Studies in Humans and Experimental Models. *Front Mol Neurosci* 2017;10:349.
- [18] Farr M, Garvey K, Bold AM, Kendall MJ, Bacon PA. Significance of the hydrogen ion concentration in synovial fluid in rheumatoid arthritis. *Clin Exp Rheumatol* 1985;3:99–104.
- [19] Firestein GS, McInnes IB. Immunopathogenesis of Rheumatoid Arthritis. *Immunity* 2017;46:183–196.
- [20] Fosang AJ, Last K, Gardiner P, Jackson DC, Brown L. Development of a cleavage-site-specific monoclonal antibody for detecting metalloproteinase-derived aggrecan fragments: detection of fragments in human synovial fluids. *Biochem J* 1995;310:337–343.
- [21] Fujii W, Kawahito Y, Nagahara H, Kukida Y, Seno T, Yamamoto A, Kohno M, Oda R, Taniguchi D, Fujiwara H, Ejima A, Kishida T, Mazda O, Ashihara E. Monocarboxylate Transporter 4, Associated With the Acidification of Synovial Fluid, Is a Novel Therapeutic Target for Inflammatory Arthritis: MCT4 IS A THERAPEUTIC TARGET FOR INFLAMMATORY ARTHRITIS. *Arthritis & Rheumatology* 2015;67:2888–2896.
- [22] Goldie I, Nachemson A. Synovial pH in rheumatoid knee-joints. I. The effect of synovectomy. *Acta Orth Scand* 1969;40:634–641.
- [23] Heiberg T, Kvien TK. Preferences for improved health examined in 1,024 patients with rheumatoid arthritis: Pain has highest priority. *Arthritis & Rheumatism* 2002;47:391–397.
- [24] Hildebrand C, Öqvist G, Brax L, Tuisku F. Anatomy of the rat knee joint and fibre composition of a major articular nerve. *Anat Rec* 1991;229:545–555.
- [25] Hunter DJ, Bierma-Zeinstra S. Osteoarthritis. *Lancet* 2019;393:1745–1759.
- [26] Inglis JJ, Notley CA, Essex D, Wilson AW, Feldmann M, Anand P, Williams R. Collagen-induced arthritis as a model of hyperalgesia: Functional and cellular analysis of the analgesic actions of tumor necrosis factor blockade. *Arth Rheum* 2007;56:4015–4023.
- [27] Izumi M, Ikeuchi M, Ji Q, Tani T. Local ASIC3 modulates pain and disease progression in a rat model of osteoarthritis. *J Biomed Sci* 2012;19:77.
- [28] Krock E, Jurczak A, Svensson CI. Pain pathogenesis in rheumatoid arthritis-what have we learned from animal models? *Pain* 2018;159 Suppl:S98–S109.
- [29] Langford LA, Schmidt RF. Afferent and efferent axons in the medial and posterior articular nerves of the cat. *Anat Rec* 1983;206:71–78.
- [30] Lewin GR, Ritter AM, Mendell LM. Nerve growth factor-induced hyperalgesia in the neonatal and adult rat. *J Neurosci* 1993;13:2136–48.

- [31] Mailhot B, Christin M, Tessandier N, Sotoudeh C, Bretheau F, Turmel R, Pellerin È, Wang F, Bories C, Joly-Beauparlant C, De Koninck Y, Droit A, Cicchetti F, Scherrer G, Boilard E, Sharif-Naeini R, Lacroix S. Neuronal interleukin-1 receptors mediate pain in chronic inflammatory diseases. *J Exp Med* 2020;217.
- [32] Marra S, Ferru-Clément R, Breuil V, Delaunay A, Christin M, Friend V, Seville S, Cognard C, Ferreira T, Roux C, Euller-Ziegler L, Noel J, Lingueglia E, Deval E. Non-acidic activation of pain-related Acid-Sensing Ion Channel 3 by lipids. *EMBO J* 2016;35:414–428.
- [33] Miller RE, Ishihara S, Tran PB, Golub SB, Last K, Miller RJ, Fosang AJ, Malfait A-M. An aggrecan fragment drives osteoarthritis pain through Toll-like receptor 2. *JCI Insight* 2018;3:e95704.
- [34] Miller RE, Tran PB, Das R, Ghoreishi-Haack N, Ren D, Miller RJ, Malfait A-M. CCR2 chemokine receptor signaling mediates pain in experimental osteoarthritis. *Proceedings of the National Academy of Sciences* 2012;109:20602–20607.
- [35] Miller RJ, Malfait A-M, Miller RE. The innate immune response as a mediator of osteoarthritis pain. *Osteoarthritis and Cartilage* 2020;28:562–571.
- [36] Qu L, Zhang P, LaMotte RH, Ma C. Neuronal Fc-gamma receptor I mediated excitatory effects of IgG immune complex on rat dorsal root ganglion neurons. *Brain, Behavior, and Immunity* 2011;25:1399–1407.
- [37] Rudjito R, Agalave NM, Farinotti AB, Lundbäck P, Szabo-Pardi T, Price TJ, Harris HE, Burton MD, Svensson CI. Sex- and cell-dependent contribution of peripheral HMGB1 and TLR4 in arthritis-induced pain. *Pain* 2020; Publish Ahead of Print. doi:10.1097/j.pain.0000000000002034.
- [38] Safiri S, Kolahi AA, Hoy D, Smith E, Bettampadi D, Mansournia MA, Almasi-Hashiani A, Ashrafi-Asgarabad A, Moradi-Lakeh M, Qorbani M, Collins G, Woolf AD, March L, Cross M. Global, regional and national burden of rheumatoid arthritis 1990–2017: a systematic analysis of the Global Burden of Disease study 2017. *Ann Rheum Dis* 2019;78:1463–1471.
- [39] Safiri S, Kolahi A-A, Smith E, Hill C, Bettampadi D, Mansournia MA, Hoy D, Ashrafi-Asgarabad A, Sepidarkish M, Almasi-Hashiani A, Collins G, Kaufman J, Qorbani M, Moradi-Lakeh M, Woolf AD, Guillemin F, March L, Cross M. Global, regional and national burden of osteoarthritis 1990-2017: a systematic analysis of the Global Burden of Disease Study 2017. *Ann Rheum Dis* 2020:annrheumdis-2019-216515.
- [40] Sagar D, Burston JJ, Hathway GJ, Woodhams SG, Pearson RG, Bennett AJ, Kendall DA, Scammell BE, Chapman V. The contribution of spinal glial cells to chronic pain behaviour in the monosodium iodoacetate model of osteoarthritic pain. *Mol Pain* 2011;7:88.
- [41] Schaible HG, Schmidt RF. Time course of mechanosensitivity changes in articular afferents during a developing experimental arthritis. *Journal of neurophysiology* 1988;60:2180–2195.
- [42] Schnitzer TJ, Easton R, Pang S, Levinson DJ, Pixton G, Viktrup L, Davignon I, Brown MT, West CR, Verburg KM. Effect of Tanezumab on Joint Pain, Physical Function, and Patient Global Assessment of Osteoarthritis Among Patients With Osteoarthritis of the Hip or Knee: A Randomized Clinical Trial. *JAMA* 2019;322:37.
- [43] Sluka KA, Rasmussen LA, Edgar MM, O'Donnell JM, Walder RY, Kolker SJ, Boyle DL, Firestein GS. Acid-Sensing Ion Channel 3 Deficiency Increases Inflammation but Decreases Pain Behavior in Murine Arthritis. *Arth Rheum* 2013;65:1194–1202.
- [44] Tran PB, Miller RE, Ishihara S, Miller RJ, Malfait AM. Spinal microglial activation in a murine surgical model of knee osteoarthritis. *Osteoarthritis and Cartilage* 2017;25:718–726.

- [45] Wigerblad G, Bas DB, Fernades-Cerqueira C, Krishnamurthy A, Nandakumar KS, Rogoz K, Kato J, Sandor K, Su J, Jimenez-Andrade JM, Finn A, Bersellini Farinotti A, Amara K, Lundberg K, Holmdahl R, Jakobsson P-J, Malmström V, Catrina AI, Klareskog L, Svensson CI. Autoantibodies to citrullinated proteins may induce joint pain independent of inflammation. *Ann Rheum Dis* 2016;75:730–738.
- [46] Zhu J, Zhen G, An S, Wang X, Wan M, Li Y, Chen Z, Guan Y, Dong X, Hu Y, Cao X. Aberrant subchondral osteoblastic metabolism modifies NaV1.8 for osteoarthritis. *eLife* 2020;9:e57656.

