

**Title**

In vivo imaging of synaptic density in neurodegenerative disorders with positron emission tomography:  
A systematic review

**Author names and affiliations**

Malouke Visser<sup>a, b</sup>, John T. O'Brien<sup>a</sup>, Elijah Mak<sup>a\*</sup>

<sup>a</sup> Department of Psychiatry, School of Clinical Medicine, Addenbrooke's Hospital, University of Cambridge, United Kingdom

<sup>b</sup> Neuropsychology and Rehabilitation Psychology, Donders Institute for Brain, Cognition and Behaviour, Radboud University, Nijmegen, The Netherlands

\*Corresponding author at: Department of Psychiatry, University of Cambridge School of Clinical Medicine, Box 189, Level E4 Cambridge Biomedical Campus, Cambridge, CB2 0SP, United Kingdom.

*E-mail address:* [fk24@cam.ac.uk](mailto:fk24@cam.ac.uk) (E. Mak).

**Author's contributions**

Malouke Visser conducted the literature searches and wrote the paper. Elijah Mak reviewed the drafts and contributed to the writing of the paper. John O'Brien reviewed the manuscript and provided critical feedback.

**Abstract**

Positron emission tomography (PET) with radiotracers that bind to synaptic vesicle glycoprotein 2A (SV2A) enables quantification of synaptic density in the living human brain. Assessing the regional distribution and severity of synaptic density loss will contribute to our understanding of the pathological processes that precede atrophy in neurodegeneration. In this systematic review, we provide a discussion of *in vivo* SV2A PET imaging research for quantitative assessment of synaptic density in various dementia conditions: amnesic Mild Cognitive Impairment and Alzheimer's disease, Frontotemporal dementia, Progressive supranuclear palsy and Corticobasal degeneration, Parkinson's disease and Dementia with Lewy bodies, Huntington's disease, and Spinocerebellar Ataxia. We discuss the main findings concerning group differences and clinical-cognitive correlations, and explore relations between SV2A PET and other markers of pathology. Additionally, we touch upon synaptic density in healthy ageing and outcomes of radiotracer validation studies. Studies were identified on PubMed and Embase between 2018 and 2023; last searched on the 3<sup>rd</sup> of July 2023. A total of 36 studies were included, comprising 5 on normal ageing, 21 clinical studies, and 10 validation studies. Extracted study characteristics were participant details, methodological aspects, and critical findings. In summary, the small but growing literature on *in vivo* SV2A PET has revealed different spatial patterns of synaptic density loss among various neurodegenerative disorders that correlate with cognitive functioning, supporting the potential role of SV2A PET imaging for differential diagnosis. SV2A PET imaging shows tremendous capability to provide novel insights into the aetiology of neurodegenerative disorders and great promise as a biomarker for synaptic density reduction. Novel directions for future synaptic density research are proposed, including (a) longitudinal imaging in larger patient cohorts of preclinical dementias, (b) multi-modal mapping of synaptic density loss onto other pathological processes, and (c) monitoring therapeutic responses and assessing drug efficacy in clinical trials.

**Keywords:** Aging; Dementia; SV2A; PET; Synaptic density; Neurodegeneration;

46 **1. Introduction**

47 Neurodegenerative disorders are a complex and pressing challenge in the field of neuroscience,  
48 demanding advanced diagnostic and monitoring techniques. A key element of neuropathology in  
49 neurodegenerative disorders is the selective loss of vulnerable populations of neurons, which plays a  
50 central role in disease progression (Dugger & Dickson, 2017). While magnetic resonance imaging  
51 (MRI) and computed tomography (CT) are capable of detecting grey matter (GM) atrophy at a  
52 macroscale level, structural imaging modalities lack the sensitivity to assess the majority of pathological  
53 processes *preceding* neuronal loss (Dugger & Dickson, 2017; Serrano et al., 2022; Topcuoglu et al.,  
54 2022b).

55 In response to this challenge, positron emission tomography (PET) has emerged as a powerful  
56 tool. PET enables more specific, *in vivo*, imaging and quantification of a wide range of physiological  
57 functions at the molecular level, including the presence of specific protein aggregates associated with  
58 neurodegeneration, such as  $\beta$ -amyloid, tau, and  $\alpha$ -synuclein. This has proven invaluable in providing  
59 insights for differential diagnostics and therapeutic monitoring (Carson et al., 2022; Dierckx et al., 2021;  
60 Herholz et al., 2007; Iaccarino et al., 2017; Topcuoglu et al., 2022a; van Waarde et al., 2021).

61 Among the multitude of valuable pathology markers, *synaptic density* has recently gained  
62 considerable attention from the scientific community (Becker et al., 2020; Cai et al., 2019; van Waarde  
63 et al., 2021). A growing body of evidence demonstrates the significance of synapse dysfunction in  
64 neurodegenerative disorders, resulting in the emerging theory that many of these conditions can be  
65 categorised as *synaptopathies* (Lepeta et al., 2016; Luo et al., 2018; Rabiner, 2018; Serrano et al., 2022).  
66 Measuring synaptic density within specific brain regions is not a novel concept. Conventional  
67 synaptophysin-based methods include stereology, immunohistochemistry, and electron microscopy  
68 (Calhoun et al., 1996). However, such methods entail *ex vivo* measurements and are thus limited by the  
69 requirement of post-mortem or surgically resected brain tissue (Finnema et al., 2016; Serrano et al.,  
70 2022).

71 Recently, the development of high-affinity PET radiopharmaceuticals targeting the synaptic  
72 vesicle protein 2A (SV2A) has enabled the ability to directly assess synaptic density in the living human  
73 brain (Finnema et al., 2016; Mercier et al., 2014; Mercier et al., 2017). SV2A is a glycoprotein present  
74 in presynaptic vesicles of both glutamatergic and GABAergic neurons throughout the central nervous  
75 system. Its ubiquitous expression has led researchers to propose SV2A as a proxy for synaptic density  
76 (Cai et al., 2019; Carson et al., 2022; Finnema et al., 2016; Serrano et al., 2022). Traditionally linked to  
77 epilepsy, SV2A serves as the target for antiepileptic drugs levetiracetam (Keppra®) and brivaracetam  
78 (Briviact®, Feng et al., 2009; Gillard et al., 2011; Lynch et al., 2004).

79 Utilising SV2A PET imaging as a potential indicator of synaptopathology, researchers can gain  
80 insights into the dynamics of synaptic density, revealing patterns of abnormality (Finnema et al., 2016;  
81 Finnema et al., 2020; Rizzoli & Betz, 2005). Consequently, these findings may pave the way for the  
82 development and assessment of new therapeutic interventions aimed at reducing synaptic toxicity.  
83 While previous narrative reviews have outlined the development, radiochemistry, and application of  
84 SV2A PET imaging agents for measuring synaptic density *in vivo* in neuropsychiatric and  
85 neurodegenerative disorders, a systematic approach in this area has yet to be undertaken (Becker et al.,  
86 2020; Cai et al., 2019; Carson et al., 2022; Martin et al., 2023; Rabiner, 2018). Here we report what is,  
87 to our knowledge, the first systematic review of SV2A PET imaging research for quantitative  
88 assessment of synaptic density in neurodegenerative disorders. Providing an objective synthesis of  
89 existing evidence, we enable a comprehensive evaluation of the current state of research, thereby aiming  
90 to illuminate the importance of SV2A PET in understanding neurodegenerative pathology.

91

92

## 93 2. Methods

94 This systematic review was reported following the guidelines of the Preferred Reporting Items for  
95 Systematic Reviews and Meta-analyses (Page et al., 2021).

### 96 2.1. Search strategy and selection criteria

97 To identify relevant studies involving PET imaging of synaptic density in dementia patients, a search  
98 was conducted in PubMed and Embase, using the following search terms: ((Positron emission  
99 tomography OR PET) AND (Synaptic density OR presynaptic density OR synaptic loss OR synaptic  
100 damage OR synaptic complexity OR UCB-J OR UCB-H OR SV2A) AND (Dementia OR  
101 Neurodegeneration OR Alzheimer disease OR Lewy body OR Parkinson disease OR Frontotemporal  
102 OR Huntington OR Progressive supranuclear palsy OR Corticobasal degeneration OR Primary  
103 tauopathies OR Mild cognitive impairment OR Normal aging OR Healthy aging OR Carriers OR  
104 Presymptomatic OR Asymptomatic OR Validation OR Test retest)). The complete search strings are  
105 reported in the supplemental materials (**Table S1**). The initial search was conducted by one author  
106 (M.V.) on the 19<sup>th</sup> of June 2023. Applied filters were language (English), peer-reviewed literature, and  
107 publication date (from January 2019). On the 3<sup>rd</sup> of July 2023, the search was extended up to January  
108 2018. Inclusion criteria encompassed studies that investigated synaptic density *in vivo* using PET  
109 imaging in the following populations: (1) healthy ageing individuals; (2) individuals with a family  
110 history of dementia; (3) carriers of dementia-associated risk genes, or; (4) patients diagnosed with a  
111 neurodegenerative disorder. Additionally, studies addressing methodological aspects of synaptic  
112 density PET imaging, specifically those aimed at validation, were also included. Excluded from  
113 considerations were case reports, reviews, letters, commentaries, and editorials. Furthermore, if  
114 multiple papers covered the same sample, the most recent study was considered the primary source to  
115 prevent duplication.

### 116 2.2. Study selection

117 After duplicate removal, a single author (M.V.) conducted the selection process by reviewing the titles  
118 and abstracts of the search results. Studies were classified as “Include” (eligible), “Maybe” (potentially  
119 eligible), or “Exclude” (not eligible) based on their relevance. Full texts were then obtained for further  
120 assessment of eligibility. Any uncertainty regarding study selection was resolved through a consensus-  
121 based discussion with a second reviewer (E.M.). For transparency, rationales for exclusion after full-  
122 text screening was documented in the supplemental materials (**Table S2**). To ensure comprehensive  
123 literature coverage, references cited in the included study reports were reviewed to identify additional  
124 relevant studies. The PRISMA flow diagram in **Figure 1** provides a visual summary of the study  
125 selection process.

### 126 2.3. Data extraction, risk of bias assessment and data synthesis

127 M.V. extracted the following study characteristics: (1) Participant details: disorder, sample size, time  
128 points (if applicable), age, and sex distribution; (2) Methodological aspects: study type, imaging  
129 specifics (i.e., modality and acquisition, radioligand used, reference region); and (3) Results: principal  
130 findings relating to group comparisons (regional and global), correlations with other pathology markers  
131 and cognitive performance. The results were presented in a table, a figure, and described in narrative  
132 form. No attempts were made to extract quantitative data for meta-analysis.

133 **3. Results**

134 The database search resulted in 200 studies. Seven studies were identified through citation screening.  
 135 Following careful evaluation, 36 key studies published between 2018 and 2023 were included in the  
 136 systematic review (see flow diagram in **Figure 1**). Included studies were focused normal ageing ( $n =$   
 137 5), individuals with cognitive impairment and various neurodegenerative conditions ( $n = 21$ ), and  
 138 validation studies on determining suitable quantification methods of SV2A expression and evaluating  
 139 the kinetic properties of radiotracers ( $n = 10$ ). Clinical studies included patients on amnesic Mild  
 140 Cognitive Impairment (aMCI) and Alzheimer's disease (AD,  $n = 9$ ), Frontotemporal dementia  
 141 behavioural variant (bvFTD,  $n = 2$ ), Progressive supranuclear palsy (Richardson's syndrome; PSP) and  
 142 CBS (Corticobasal syndrome without Alzheimer's pathology, otherwise referred to as Corticobasal  
 143 degeneration; CBD,  $n = 4$ ), Parkinson's disease (PD) and dementia with Lewy bodies (DLB,  $n = 4$ ),  
 144 Huntington's disease (HD,  $n = 1$ ), and Spinocerebellar Ataxia Type 3 (SCA3,  $n = 1$ ). A summary of the  
 145 clinical study characteristics and principal findings, grouped by publication date and neurodegenerative  
 146 disorder, is described in **Table 1**. Key findings will be interpreted and discussed in further detail in  
 147 **Section 4.3**.

148 Clinical study methods exhibited some notable commonalities and differences. Approximately  
 149 half of the studies ( $n = 10$ ) included a sample size of 20 or more subjects with a neurodegenerative  
 150 disorder. The majority of studies ( $n = 16$ ) exclusively used cross-sectional data while there were five  
 151 longitudinal studies across aMCI ( $n = 1$ ), AD ( $n = 1$ ), PSP and CBS ( $n = 1$ ), and PD ( $n = 2$ ).

152 All studies integrated T1-weighted MRI scanning to control for cases with anatomical  
 153 abnormalities, facilitate co-registration with PET data and define regions of interest if applicable. PET  
 154 imaging data was largely captured dynamically ( $n = 16$ ), but studies utilised different types of PET  
 155 machines (PET ( $n = 7$ ), PET-CT ( $n = 7$ ), or PET-MR scanners ( $n = 7$ )). The predominant tracer choice  
 156 was [ $^{11}\text{C}$ ]UCB-J ( $n = 17$ ), while a smaller number of studies selected relatively novel tracers [ $^{18}\text{F}$ ]UCB-  
 157 H ( $n = 2$ ) and [ $^{18}\text{F}$ ]SynVesT-1 ( $n = 2$ ). Moreover, a notable diversity was observed in the methods  
 158 employed to quantify the PET binding signal, with nondisplaceable binding potential ( $\text{BP}_{\text{ND}}$ ) being the  
 159 prevailing metric in  $n = 8$  studies. Followed by standardised uptake value ratio (SUVR  $n = 3$ , SUVR-1  
 160  $n = 3$ ), distribution volume ratio ( $\text{DVR}_{\text{CBL}} n = 3$ ,  $\text{DVR}_{\text{SO}} n = 1$ ), and volume of distribution ( $V^{\text{T}}$ ,  $n = 3$ ).  
 161 A limited number of studies ( $n = 4$ ) employed arterial cannulation to obtain the arterial input function  
 162 (AIF). In contrast, most studies ( $n = 17$ ) opted for non-invasive alternatives for arterial sampling. This  
 163 encompassed the application of simplified reference tissue modelling ( $n = 9$ ), the calculation of SUVR  
 164 ( $n = 6$ ), and the implementation of image-derived input functions (IDIF,  $n = 2$ ; both in the [ $^{18}\text{F}$ ]UCB-H  
 165 tracer studies).

166 With regards to statistical analysis, a large number of studies ensured that experimental groups  
 167 were matched for age ( $n = 18$ ) and sex ( $n = 15$ ), however relatively few described efforts to match for  
 168 education ( $n = 9$ ). Two studies did not make direct comparisons between groups (Coomans et al., 2021;  
 169 Mecca, O'Dell, et al., 2022). To correct for multiplicity, studies mostly applied FDR ( $n = 9$ ), but FWE  
 170 ( $n = 4$ ) and Bonferroni ( $n = 3$ ) correction methods were also conducted. Additionally, two studies did  
 171 not report which correction method was used (Chen et al., 2018; Matuskey et al., 2020), one study  
 172 employed both FDR and FWE (Malpetti et al., 2023), and two studies exclusively reported uncorrected  
 173 results (Coomans et al., 2021; Venkataraman et al., 2022). All but two study corrected for Partial  
 174 Volume Effects (PVEs, Coomans et al., 2021; Wilson et al., 2020). Applied voxel-based Partial Volume  
 175 Correction (PVC) methods were the Muller-Gartner method (MG,  $n = 5$ ), the Iterative Yang algorithm  
 176 (IY,  $n = 5$ ), and the region-based voxel-wise correction (RBV,  $n = 4$ ). The most frequently used region-  
 177 based method was the geometric transfer matrix (GTM,  $n = 4$ ). Malpetti et al. (2023) applied both IY  
 178 and GTM correction methods for voxel-wise and region of interest (ROI) analysis respectively.

179 A large proportion of studies (94.74%) that conducted direct statistical group comparisons ( $n =$   
 180 19) reported decreased synaptic density in one or more brain areas among individuals with

181 neurodegenerative disorders compared to healthy controls (HC,  $n = 18$ , **Figure 2A**). Furthermore, most  
182 studies (73.33%) that inquired into the relation between synaptic density and cognitive functioning ( $n$   
183 = 15) reported associations between synaptic loss and global or domain-specific cognitive decline ( $n =$   
184 11, **Figure 2B**). These significant correlations were evident in studies focusing on AD and aMCI ( $n =$   
185 6), Frontotemporal lobar degeneration pathologies (FTLD,  $n = 4$ ), and Lewy body diseases ( $n = 1$ ). See  
186 supplemental materials for a detailed visualisation of these results (**Figure S3**).

187 **Figure 1.** PRISMA flow diagram of the literature search (**filename = Figure.1.colourised\_M.Visser**)  
188 *Notes.* Adapted from Page et al. (2021).

189 **Figure 2.** Percentage of clinical studies showing group differences in synaptic density and correlations  
190 with cognitive functioning (**filename = Figure.2.colourised\_M.Visser**) *Notes.* “Decreased synaptic  
191 density” indicates the presence of statistically significant group differences (adjusted for multiple  
192 comparisons) in one or more brain regions between diseased subjects and controls. Longitudinal studies  
193 were included if they identified cross-sectional differences at baseline, follow-up, or both. “Cognitive  
194 correlations” indicate statistically significant associations (adjusted for multiple comparisons) between  
195 synaptic density in one or more brain regions and neuropsychological tests measuring global or domain-  
196 specific cognitive functioning in diseased subjects, excluding motor functioning. Longitudinal studies  
197 were included if associations between changes in synaptic density over time and cognitive performance  
198 were found.

199

## 200 4. Discussion

201 The primary objective of this systematic review was to offer a thorough summary of current research  
202 on *in vivo* SV2A PET imaging in neurodegenerative disorders. Discussing the novel and rapidly  
203 developing area of *in vivo* SV2A PET imaging, we begin by reviewing the outcomes of validation  
204 studies. Afterwards, we address synaptic density in the context of normal ageing and extend our scope  
205 by encompassing its manifestations in various neurodegenerative disorders: aMCI, AD, FTLN  
206 pathologies, Lewy body diseases, HD, and SCA3. Following this, the relation of *in vivo* synaptic density  
207 loss to cognitive decline, as well as other markers of pathologies is explored.

### 208 4.1. Validation studies

#### 209 4.1.1. Tracer kinetic modelling

210 A range of kinetic models have been investigated to identify the optimal kinetic analysis model for *in*  
211 *vivo* SV2A PET. In current literature, full kinetic modelling with arterial blood sampling confirmed the  
212 1-Tissue-Compartment (1TC) model as most suitable to describe cerebral [<sup>11</sup>C]UCB-J and  
213 [<sup>18</sup>F]SynVesT-1 tracer kinetics (Finnema et al., 2018; Koole et al., 2019; Li et al., 2021; Tuncel et al.,  
214 2021). Given the strong correlation between the Multilinear Analysis 1 (MA1) and 1TC-derived  $V_T$   
215 estimates, MA1 may also be considered appropriate (Mansur et al., 2020; Naganawa et al., 2021).  
216 Furthermore, in various studies, Akaike Information Criterion and F-test preferred the 2-Tissue-  
217 Compartment (2TC) model over 1TC, however, the 2TC faced challenges in reliably estimating kinetic  
218 parameters (Mansur et al., 2020; Naganawa et al., 2021; Tuncel et al., 2021). Results were inconclusive  
219 regarding the additional inclusion of a blood volume fraction as a parameter (1T2k\_  $V_B$ ) into this kinetic  
220 evaluation. Finnema et al. (2018) reported 1T2k\_  $V_B$  inclusion did not affect regional  $V_T$  values, causing  
221 Rossano et al. (2020) to obtain from using it. Others did include the 1T2k\_  $V_B$  metric (Koole et al., 2019;  
222 Mansur et al., 2020; Tuncel et al., 2021), with Tuncel et al. (2021) and Mansur et al. (2020) suggesting  
223 it enhances the description of [<sup>11</sup>C]UCB-J kinetics.

#### 224 4.1.2. Reference tissue modelling

225 To make PET radioligands more suitable for clinical use, the preference lies in utilising practical, non-  
226 invasive alternative approaches to arterial blood sampling. One approach is the use of reference tissue  
227 models, which allow for the quantification of  $BP_{ND}$  (Zanderigo et al., 2013). Various studies have  
228 validated different parametric methods for quantifying SV2A binding using subcortical white matter  
229 (WM) as reference tissue (i.e., centrum semi-ovale; SO). These included the Simplified Reference  
230 Tissue Model (SRTM), static SUVR, Multilinear Reference Tissue Model (MRTM), and reference  
231 Logan Graphical Analysis (rLGA). Across studies, the SRTM<sub>SO</sub> was considered the most appropriate  
232 choice for [<sup>11</sup>C]UCB-J quantification. STRM2 corresponds best to plasma input-derived DVR, although  
233 signal underestimation was observed (Koole et al., 2019; Mertens et al., 2020; Tuncel et al., 2021;  
234 Tuncel et al., 2022). Additionally, a SUVR<sub>SO</sub> approach was also identified as a valuable alternative for  
235 full kinetic modelling or STRM, particularly since the precision of the latter decreases when scanning  
236 time is <60 min, limiting clinical applicability (Koole et al., 2019; Mertens et al., 2020).

#### 237 4.1.3. Reference tissue selection

238 The SO has been validated as a reference region for the quantification of SV2A density. Comparing  
239 baseline and post-SV2A-drug (Padsevoni) scanning [<sup>11</sup>C]UCB-J, no significant changes in the SO  
240 distribution volume (1TC-derived) were identified. Meanwhile, regional  $V_T$  values were significantly  
241 reduced in other regions, suggesting negligible target expression in the SO (Koole et al., 2019). Further  
242 supporting its potential as reference tissue, Mansur et al. (2020) demonstrated [<sup>11</sup>C]UCB-J  $V_T$  estimates  
243 to be approximately 60% lower in the SO. Tuncel et al. (2021) additionally found no significant  
244 difference in SO  $V_T$ 's between AD patients and HC.

245 Ideally, there should be no specific or displaceable tracer uptake in a reference region. In  
 246 response to Finnema et al. (2016), who indicated the presence of displaceable activity in the SO,  
 247 Rossano et al. (2020) further carefully characterised SO [<sup>11</sup>C]UCB-J binding. Even when they  
 248 minimised the effects of GM matter spill-in, partial volume, and low-intensity WM biases, displaceable  
 249 SO [<sup>11</sup>C]UCB-J uptake was not fully eliminated. In particular, SO V<sub>T</sub> overestimates GM V<sub>ND</sub> uptake.  
 250 Consequently, BP<sub>ND</sub> is negatively biased if SO is used as a reference. Nevertheless, SO V<sub>T</sub> and GM V<sub>ND</sub>  
 251 estimates are highly intercorrelated and potential differences in [<sup>11</sup>C]UCB-J behaviour between GM and  
 252 WM tissue types may be consistent within and between groups (Rossano et al., 2020; Tuncel et al.,  
 253 2022). Therefore, the SO region is still thought to be a suitable reference region for [<sup>11</sup>C]UCB-J.

254 The current review did not include studies validating the use of a cerebellar (CBL) reference  
 255 region, as these were excluded due to overlapping samples with more recent publications (see **Table**  
 256 **S3**). Nonetheless, given its relevance, validation of conversion from SO to CBL reference regions in  
 257 [<sup>11</sup>C]UCB-J PET, as conducted by Mecca et al. (2020) and O'Dell et al. (2021), will be addressed briefly.  
 258 Overall, values of DVR<sub>CBL</sub> demonstrated notably lower variability than DVR<sub>CS</sub> and showed a  
 259 comparable level of correlation with DVR values obtained using the 1TC model and metabolite-  
 260 corrected arterial plasma curves (Mecca et al., 2020; O'Dell et al., 2021). More validation studies  
 261 assessing the potential of CBL as a reference region are warranted.

#### 262 4.1.4. Test-retest reliability

263 For [<sup>11</sup>C]UCB-J V<sub>T</sub>, absolute test–retest (TRT) reproducibility ranges between 3–9% across regions in  
 264 healthy subjects, which is exceptionally good compared to TRT of other tracers previously reported  
 265 (Finnema et al., 2018). In AD patients, a mean 28-day TRT of <15% was identified for V<sub>T</sub>, plasma  
 266 input-derived DVR and SRTM BP<sub>ND</sub>. This suggests that [<sup>11</sup>C]UCB-J can be used to quantify the effect  
 267 of therapeutical interventions on SV2A if the effect size of the intervention is higher than 15% (Tuncel  
 268 et al., 2021). Tuncel et al. (2022) further evaluated TRT for whole-brain analysis and found that  
 269 reproducibility was much better for HC compared to patients with AD. However, larger TRT variability  
 270 in patients is to be expected given synaptic density reductions depend on disease severity and GM  
 271 atrophy, which may vary per individual (Tuncel et al., 2022). In comparison to [<sup>11</sup>C]UCB-J, absolute  
 272 TRT reproducibility for [<sup>18</sup>F]SynVesT-1 (performed 7 ± 7 days apart) was similar for V<sub>T</sub> and slightly  
 273 less for BP<sub>ND</sub> across regions, with values of <9% (Li et al., 2021). Overall, both same-day and 28-day  
 274 TRT reproducibility of [<sup>11</sup>C]UCB-J and [<sup>18</sup>F]SynVesT-1 V<sub>T</sub> and BP<sub>ND</sub> are found to be excellent  
 275 (Finnema et al., 2018; Li et al., 2021; Tuncel et al., 2021; Tuncel et al., 2022).

#### 276 4.2. Synaptic density in cognitively normal individuals

##### 277 4.2.1. Preservation of synaptic density during healthy ageing

278 Accurate interpretation of *in vivo* SV2A PET imaging studies in dementia necessitates careful  
 279 consideration of synaptic density in healthy individuals as well as the potential impact of demographic  
 280 factors. Michiels et al. (2021) were the first to investigate the effects of ageing and sex on [<sup>11</sup>C]UCB-J  
 281 binding *in vivo*. Their study of 78 cognitively normal subjects aged 18-85 years revealed remarkable  
 282 stability of synaptic density with advancing age, except for a slight reduction in the caudate nucleus  
 283 (1.7% per decade), possibly linked to ventricle enlargement. Additionally, no substantial sex-based  
 284 differences in synaptic density or age-sex interaction were observed (Michiels et al., 2021). Andersen  
 285 et al. (2022) expanded upon these findings with a study involving 26 healthy subjects divided into two  
 286 age groups (20-30 and 65-85). In addition to assessing synaptic density using [<sup>11</sup>C]UCB-J, cerebral  
 287 metabolism was examined using [<sup>18</sup>F]FDG. Results reaffirmed the findings of Michiels et al. (2021),  
 288 providing further evidence that, although synaptic loss contributes to the overall loss of GM volume  
 289 during ageing, the density of the synapses per volume of GM integrity remains preserved in the healthy  
 290 ageing brain. In other words, volume loss is proportional to synaptic loss with ageing, resulting in stable  
 291 synaptic density (Andersen et al., 2022). Additionally, a widespread reduction of [<sup>18</sup>F]FDG uptake in  
 292 the older age group was observed, aligning with the findings of numerous other studies (Chetelat et al.,

293 2013; Fujimoto et al., 2008; Hsieh et al., 2012; Yoshizawa et al., 2014). Moreover, regional differences  
 294 in relative [ $^{18}\text{F}$ ]FDG vs. [ $^{11}\text{C}$ ]UCB-J uptake were identified, suggesting “some synapses are on average  
 295 more hungry than others”, given their higher relative metabolic demands (Andersen et al., 2022, p. 56).  
 296 These findings align with an earlier multi-tracer study by van Aalst et al. (2021) in 20 young healthy  
 297 female subjects. Both studies observed higher relative [ $^{11}\text{C}$ ]UCB-J uptake in the temporal lobe and  
 298 higher relative [ $^{18}\text{F}$ ]FDG uptake in frontal areas (Andersen et al., 2022; van Aalst et al., 2021). There  
 299 thus seems to exist a spatially variant decoupling of relative synaptic density and metabolic activity in  
 300 healthy individuals. Interestingly, this relationship may be slightly affected by age, especially in the  
 301 frontal lobe, where lower glucose consumption per synapse was observed in the older group (2%)  
 302 compared to the younger group (5%). However, the limited sample size in Andersen et al.'s (2022) study  
 303 cautions against making definitive conclusions from this discovery. Overall, studies collectively support  
 304 the notion that synaptic density remains relatively preserved during healthy ageing, despite age-related  
 305 changes in cerebral metabolism and grey matter volume.

#### 306 4.2.2. *Exploring the relationship between synaptic density and resting-state network functioning*

307 Synaptic density has not only been linked to local synaptic activity, but also to the healthy brain's  
 308 functional organisation, that is, resting-state network (RSN) activity (Fang et al., 2021; Fang et al.,  
 309 2023). Fang et al. (2021) applied independent component analysis (ICA) to [ $^{11}\text{C}$ ]UCB-J  $V_T$  data and  
 310 reliably extracted multiple source-based patterns (i.e., networks) of coherent synaptic density  
 311 variability. Based on visual comparison, Fang and colleagues argue that the spatial organisation of  
 312 synaptic density network variability shows some similarity with that of canonical RSNs, but appears to  
 313 be more consistent with GM and glucose metabolism (Fang et al., 2021). Subsequently, in their 2023  
 314 study, (Fang et al.) conducted a quantitative investigation into this relationship. A robust correlation was  
 315 identified between functional amplitudes in the anterior default-mode network and networks of synaptic  
 316 density that are both anatomically connected (e.g., the medial prefrontal cortex), and functionally  
 317 associated (e.g., the striatum). Contrary to expectations, no other strong relations were found between  
 318 networks of synaptic density and spatially overlapping or functionally associated RSNs of interest,  
 319 implying the degree to which synaptic density underlies RSN amplitudes is multifactorial and requires  
 320 further investigation (Fang et al., 2021; Fang et al., 2023).

321

#### 322 4.3. *Application of SV2A PET imaging in the investigation of neurodegenerative disorders*

##### 323 4.3.1. *Pairwise comparisons*

##### 324 4.3.1.1. *Alzheimer's disease*

325 Chen et al. (2018) were the first to study *in vivo* SV2A PET in AD and aMCI. They demonstrated lower  
 326 [ $^{11}\text{C}$ ]UCB-J  $\text{BP}_{\text{ND}}$  in the hippocampus (-41%) of 9 mild AD/aMCI patients compared to 11 HC  
 327 (SRTM $_{2\text{SO}}$ -derived). Exploratory voxel-wise analysis suggested an additional significant binding  
 328 reduction in the entorhinal cortex; however, this result was uncorrected for multiplicity and did not  
 329 withstand PVC. In a larger sample size, using an SV2A radiotracer with a longer half-life ([ $^{18}\text{F}$ ]UCB-  
 330 H;  $T_{1/2} = 110$  min), Bastin et al. (2020) confirmed the findings of Chen et al. (2018) and revealed  
 331 significant synaptic density reduction ( $V_T$ ) in the right anterior hippocampus (-26.9%) in AD/aMCI  
 332 patients, which extended to the entorhinal cortex. Subsequently, based on earlier studies by Mecca et  
 333 al. (2020) and O'Dell et al. (2021) -not included in the current review due to overlapping samples-, Chen  
 334 et al. (2021) adjusted their methodology. When regional distribution volume ratios were computed  
 335 using the cerebellum as an alternative reference region, widespread synaptic density loss was observed  
 336 in medial temporal and thalamic regions, as well as some small between-group differences in  
 337 susceptible neocortical regions (-7.6% to -24.8%, Chen et al., 2021). In a related study, Mecca, Chen,  
 338 et al. (2022) reported reduced hippocampal [ $^{11}\text{C}$ ]UCB-J binding ( $\text{DVR}_{\text{CBL}}$ ) in AD/aMCI, but were  
 339 unable to statistically confirm group differences. This null-finding might be attributed to the fact that  
 340 Chen et al. (2021) directly derived DVRs for the ROIs from ITC parameters using AIF, rather than  
 341 using the reference-region-based analysis employed by Mecca, Chen, et al. (2022). Furthermore, in  
 342 aMCI exclusively, cross-sectional evaluation at baseline and follow-up revealed a significant decrease

343 in SV2A binding in the mesotemporal cortex, which had progressed to the entire mesotemporal, inferior  
344 frontal, precuneus and temporo-occipital secondary cortex two years later (Vanderlinden et al., 2022).  
345 In AD patients, Venkataraman et al. (2022) also demonstrated a focal AD-related decrease in synaptic  
346 binding, largest in the caudate (-25%), hippocampus (-24%), and thalamus (-19%). Most recently, a  
347 larger cohort study utilising a difluoro-analogue of UCB-J, reported reduced binding of [<sup>18</sup>F]SynVesT-  
348 1 in the hippocampus and bilateral cortex of AD patients. In comparison to aMCI, AD patients showed  
349 reduced synaptic density in the middle frontal gyrus and right insular cortex (Zhang et al., 2023).  
350 Together, these findings indicate the medial temporal lobe is the most robust site of synaptic density  
351 loss across the continuum of AD, evident across a diverse range of methodologies and radiotracers.  
352 Evidence for reduced synaptic density in association cortices is somewhat inconsistent, possibly due to  
353 limited statistical power and the inclusion of predominantly early-stage AD patients.

#### 354 4.3.1.2. Frontotemporal lobar degeneration pathologies

355 Initial *in vivo* evidence of synaptic loss in FTLD pathologies came indirectly through a case report by  
356 Malpetti et al. (2021) on risk-gene carriers. Subsequently, Salmon et al. (2021) conducted the first  
357 exploratory research in 12 bvFTD (i.e., behavioural variant) and 12 HC participants. A trend-level  
358 reduction of [<sup>18</sup>F]UCB-H V<sub>T</sub> in the right parahippocampal region (-41%) in bvFTD compared to HC  
359 was reported, but no frontal synaptic loss or difference between bvFTD and AD was identified (Salmon  
360 et al., 2021). A subsequent study employed a radiotracer with higher specific binding, a kinetic analysis  
361 with more sensitivity to synaptic changes ([<sup>11</sup>C]UCB-J BP<sub>ND</sub>), as well as a sample comprising patients  
362 with longer symptom duration and lower cognitive performance (Malpetti et al., 2023). Reportedly,  
363 synaptic loss was severe, with binding reduction being most pronounced in frontotemporal, insular, and  
364 cingulate regions. Moreover, the effect sizes and regional extent of disease-related synaptic density loss  
365 were notably larger than those of GM atrophy (Malpetti et al., 2023). In PSP and CBS, a similar pattern  
366 was observed, with synaptic density demonstrating a more pronounced and widespread degree of  
367 reduction compared to the extent of GM atrophy (Holland et al., 2023; Holland et al., 2020; Holland et  
368 al., 2022; Mak et al., 2021). It is worth noting that PSP/CBS studies were conducted using a highly  
369 similar methodology, allowing for more reliable and comparable results. In patient cohorts compared  
370 to HC, reduced [<sup>11</sup>C]UCB-J BP<sub>ND</sub> was consistently reported in all cortical lobes, as well as in the  
371 cingulate, hippocampus, insula, amygdala, and subcortical structures across studies. Severe synaptic  
372 density loss was observed in similar regions in both PSP and CBS (i.e., medulla and caudate nucleus),  
373 but also in distinct regions between the two conditions (no statistical testing). Overall, frontotemporal  
374 lobar degeneration pathologies are marked by widespread, severe synaptic density loss in both cortical  
375 and subcortical regions, including regions affected in later stages of disease and non-atrophied areas.

#### 376 4.3.1.3. Lewy body diseases

377 Matuskey et al. (2020) provided the first evidence of synaptic loss in PD. In 12 moderate to advanced  
378 PD patients compared to 12 HC, reduced [<sup>11</sup>C]UCB-J BP<sub>ND</sub> was reported in PD-related brainstem nuclei,  
379 particularly in the substantia nigra (SN, -45%), red nucleus (-31%), locus coeruleus (-17%), as well  
380 as in some cortical areas, such as the parahippocampal gyrus (-12%). In a pilot study with 12 very early  
381 drug-naïve PD patients, Wilson et al. (2020) corroborated this finding and observed reduced lower  
382 synaptic density ([<sup>11</sup>C]UCB-J V<sub>T</sub>) in primary PD-related brain regions as well, although less pronounced  
383 in the SN (-7%). Note that effects of tissue atrophy could not be ruled out as no PVC was performed  
384 (Wilson et al., 2020). A larger cohort of 27 early PD and 18 HC participants showed [<sup>11</sup>C]UCB-J was  
385 significant in the SN (-15%), consistent with baseline results (Delva et al., 2020), and at trend-level in  
386 striatal areas (-6% to -7%, Delva, Van Laere, et al., 2022). Furthermore, in a mixed sample of patients  
387 with PD and dementia (PDD) and DLB, Andersen et al. (2023) demonstrated clear reductions in  
388 synaptic density, as measured with [<sup>11</sup>C]UCB-J SUVR-1, across most cortical regions (-21% to -46%).  
389 Collectively, these findings suggest that synaptic density loss occurs early in the pathophysiology of

390 PD, beginning in PD-related brainstem nuclei like the substantia nigra, and extending to the cortex in  
391 more advanced stages that come with cognitive impairment.

#### 392 4.3.1.4. *Other neurodegenerative disorders*

393 At present, Delva, Michiels, et al. (2022) is the only study providing *in vivo* evidence of early,  
394 multifocal, synaptic density loss in HD with [<sup>11</sup>C]UCB-J PET (Rub et al., 2016; Vonsattel et al., 2011).  
395 HD is caused by CAG expansion in the *IT15* gene, leading to the production of a polyglutamine (polyQ)  
396 stretch in the huntingtin protein. In premanifest HD, synaptic loss was identified in putamen (-19%)  
397 and caudate (-16%) only. As for early manifest HD, further extra striatal decreases in SV2A binding  
398 were observed in the pallidum (-30%), as well as cortically (-11% to -12%). Unexpectedly, SV2A loss  
399 in the cerebellum (-14%) was discovered, providing support for cerebellar involvement in the early  
400 stages of symptomatic HD (Delva, Michiels, et al., 2022). Recently, an SV2A PET study was conducted  
401 on another polyQ-related disorder, namely Spinocerebellar ataxia (Machado-Joseph disease; SCA3,  
402 Chen et al., 2023). SCA3 is instigated by CAG repeat expansion in the *ATXN3*, encoding for a polyQ  
403 stretch in the ataxin-3 protein. Compared to HC, preataxic SCA3 patients exhibited [<sup>18</sup>F]SynVesT-1  
404 SUVR reductions only in the vermis (-6%). In ataxic SCA3 patients, more significant synaptic loss was  
405 observed in the vermis (-16%), extending to extra-cerebellar regions, encompassing the brainstem,  
406 caudate, putamen, and occipital lobe (-8% to -12%). Collectively, HD and SCA3 studies indicate  
407 significant reductions in synaptic density, evident even in the absence of severe symptoms, with HD  
408 showing pronounced loss in the striatum and SCA3 demonstrating reduction specifically in the vermis.

#### 409 4.3.2. *Longitudinal SV2A PET imaging in (pre)clinical dementia*

410 To date, longitudinal research has been conducted in aMCI (Vanderlinden et al., 2022), AD  
411 (Venkataraman et al., 2022), PD (Delva, Van Laere, et al., 2022; Wilson et al., 2020) and PSP and CBS  
412 (Holland et al., 2023). Although evidence for progressive *in vivo* synaptic loss is limited, variations in  
413 the pace of longitudinal decrease of synaptic density appear to exist across different neurodegenerative  
414 conditions. Along the Alzheimer's continuum, synapse density loss may show a more progressive  
415 pattern in patients at earlier stages of the disease spectrum. In aMCI patients exclusively, Vanderlinden  
416 et al. (2022), revealed longitudinal disparities in the left superior frontal cortex over a two-year follow-  
417 up period. Meanwhile, when solely investigating AD patients, no significant progressive loss in DVR<sub>SO</sub>  
418 was identified 12 to 18 months later (Venkataraman et al., 2022). Note that Vanderlinden et al. (2022)  
419 used SUVR as the primary outcome measure, which is potentially biased due to changes in cerebral  
420 blood flow (CBF) and may be dependent on the extent of the underlying amyloid- $\beta$  burden (Heeman et  
421 al., 2022). In comparison to AD, the rate of SV2A decline in early-stage PD appears to be relatively  
422 slow. In Wilson et al. (2020), 8 PD patients underwent a longitudinal [<sup>11</sup>C]UCB-J PET scan at a 1-year  
423 interval with no significant changes detected. The caudate exhibited the most notable reduction in SV2A  
424 levels; however, this observation did not attain statistical significance (Wilson et al., 2020). A second  
425 study by Delva, Van Laere, et al. (2022) attained a longer scan interval and larger sample size.  
426 Nevertheless, no [<sup>11</sup>C]UCB-J SUVR-1 longitudinal changes were identified, suggesting synaptic  
427 density might not be a sensitive biomarker during the early phases of Lewy body diseases. The opposite  
428 seems to be true for PSP and CBS, providing promising results. Rapid longitudinal progression in  
429 [<sup>11</sup>C]UCB-J BP<sub>ND</sub> has been found in the frontal lobe (-3%), caudate nucleus (-4%), and left pallidum (-  
430 4%, trend level) after one year (Holland et al., 2023). Regardless, the consensus among these studies  
431 remains that further longitudinal investigations are necessary to ascertain whether SV2A PET possesses  
432 the capacity to serve as a clinical biomarker for monitoring the advancement of neurodegenerative  
433 disorders.

#### 434 4.3.3. *Synaptic density and cognitive functioning*

Significant correlations were apparent between reduced synaptic density and overall cognitive decline  
in studies investigating patients with aMCI/AD (Bastin et al., 2020; Chen et al., 2018; Mecca, O'Dell,

et al., 2022; Zhang et al., 2023), FTLN (Holland et al., 2023; Holland et al., 2020), and DLB (Andersen et al., 2023). Generally, reported correlations were specific to disease-related regions, involving areas such as the hippocampus regions in aMCI/AD, as well as frontal and cingulate regions in FTLN pathologies. Notably, along the continuum of Alzheimer's disease, synaptic density is a significant predictor of cognitive performance across all domains, including language (Venkataraman et al., 2022), verbal and episodic memory (Bastin et al., 2020; Chen et al., 2018; Vanderlinden et al., 2022), and executive functioning, processing speed, and visuospatial ability (Mecca, O'Dell, et al., 2022). In contrast, within Lewy body pathologies, the connection between synaptic density and domain-specific cognitive performance is notably weaker and limited. Such correlations were only observed within a specific subset of cases involving late-stage PD and DLB (Andersen et al., 2023). The exploration of whether synaptic density is associated with domain-specific cognition in the context of other neurodegenerative disorders remains an unexplored area.

Collectively, studies suggest synaptic density contributes significantly to cognitive impairment, and the application of SV2A PET imaging allows for precise mapping of behaviour and synaptic changes. Nevertheless, the extent to which synaptic density prevails over other metrics in predicting cognitive impairment remains uncertain, hinging on the pathological characteristics of each disease and their sequential progression. Notably, in cases where atrophy is minimal or absent, synaptic loss emerges as a critical mediator of cognitive decline (Malpetti et al., 2022; Mecca, O'Dell, et al., 2022). Conversely, Coomans et al. (2021) report cognitive functioning to be more strongly associated with tau build-up instead of synaptic density. Presently, cognitive outcome measures seem to primarily serve exploratory purposes. There is a need for future studies on more in-depth neuropsychological assessments sensitive to subtle focal brain changes occurring in preclinical dementia (Ritchie et al., 2017).

#### *4.3.4. Relation of in vivo synaptic density loss to other markers of pathologies*

##### *4.3.4.1. Synaptic density and hypometabolism*

Several SV2A studies have employed [<sup>18</sup>F]FDG to assess the relationship between synaptic density and reduced regional cerebral glucose consumption, indicative of impaired metabolic functionality (Dierckx et al., 2021). FDG PET detects not only cerebral hypometabolism but is also reputed to be an indirect measure of neuronal integrity (Carapelle et al., 2020). Consequently, alterations in synaptic density may lead to a decrease in cerebral energy requirements, and conversely, changes in energy demand might influence synaptic density. Exploratory analysis by Bastin et al. (2020) indicated an absence of correlation between AD-related loss of synaptic density in the hippocampus and posterior cingulate cortex and hypometabolism observed less than four years prior. This could be attributed to either a time-related progression in pathology or the capture of distinct pathological processes by the two radiotracers. Related studies acquired [<sup>18</sup>F]UCB-H and [<sup>18</sup>F]FDG PET imaging within a shorter period. In general, studies revealed positive correlations between hypometabolism and reduced synaptic density (Andersen et al., 2023; Chen et al., 2021; Delva, Michiels, et al., 2022). Chen et al. (2021) and Andersen et al. (2023) both report a similar magnitude of reduced [<sup>11</sup>C]UCB-J binding and [<sup>18</sup>F]FDG metabolism in the medial temporal lobe. Moreover, the magnitude of hypometabolism was significantly greater relative to synaptic density loss in neocortical regions. Additionally, inter-tracer correlations in medial temporal regions were notably higher compared to neocortical regions (Andersen et al., 2023; Chen et al., 2021). Overall, this suggests a regional decoupling of synaptic density and metabolic activity may be present in both healthy (see **section 4.2**) and neurodegenerative disease states. Nevertheless, the extent of cortical hypometabolism does not surpass synaptic loss in every neurodegenerative condition. Results of Delva, Michiels, et al. (2022) suggest that in HD, cortical [<sup>11</sup>C]UCB-J PET uptake is more extensive, compared to [<sup>18</sup>F]FDG uptake.

##### *4.3.4.2. Synaptic density and tau*

*In vivo* evidence indicating a link between tau pathology and synaptic density loss is prominent. Across subjects and ROIs, higher regional tau deposition, as measured with [<sup>18</sup>F]flortaucipir and [<sup>18</sup>F]MK-6240, is associated with reduced synaptic density (Coomans et al., 2021; Mecca et al., 2020; Vanderlinden et al., 2022). This relation is particularly apparent for subjects with substantial tau pathology in neocortical areas (Coomans et al., 2021), and found to be stronger compared to the inverse associations evident between regional tau and GM volume (Mecca et al., 2020). Furthermore, spatial overlap between higher [<sup>18</sup>F]flortaucipir and lower [<sup>11</sup>C]UCB-J binding is substantial (Coomans et al., 2021; Mecca et al., 2020). In line with the baseline study of Vanderlinden et al., 2022, (Vanhaute et al., 2020), the spatial pattern of tau burden seems to be slightly more widespread and pronounced compared to that of synaptic loss (Coomans et al., 2021). Additionally, a time delay between both biomarkers was found, suggesting the spatial progression of synaptic loss may follow that of tau pathology (Vanderlinden et al., 2022). In regions exhibiting elevated tau pathology, a decrease in synaptic density emerges with disease progression (Holland et al., 2022). Cumulatively, existing literature aligns with the hypothesis that tau is to precede and potentially drive the loss of synaptic function (Jack et al., 2018; Wu et al., 2021).

#### *4.3.4.3. Synaptic density and other metrics*

Instances of disease-related reduced uptake of [<sup>11</sup>C]UCB-J have also been associated with less commonly employed metrics of pathology. For example, synaptic density loss has been linked to modified synaptic functionality, suggestive of slowing of oscillatory activity (Coomans et al., 2021), and impaired functional and related structural connectivity underlying cognitive impairment (Zhang et al., 2023). Furthermore, Mak et al. (2021) have revealed a tight coupling between synaptic density loss and altered postsynaptic dendritic density and complexity. This was quantified using the Neurite Density Index and the Orientation Dispersion Index (NODDI ODI), which is an emerging diffusion-weighted MRI-based biomarker to quantify the density of neurites and their orientational complexity (Mak et al., 2021; Zhang et al., 2012). The varied effects of synaptic density loss on neural activity, both in terms of functional communication and physical wiring, along with concurrent microstructural alterations collectively point to the complexity of the synaptic landscape. Further cross-validation of more advanced imaging markers with synaptic density is warranted.

#### *4.4. Challenges to consider when interpreting SV2A PET imaging results*

Here, we briefly discuss challenges posed by potential confounding factors and heterogeneity in study methodologies. Cerebral atrophic changes may lead to signal loss due to PVEs. A decrease in apparent radiotracer uptake may thus not exclusively indicate decreased synaptic density, but could also be influenced by a concurrent disease-related loss of GM tissue (Drzezga et al., 2014). To disentangle atrophy from SV2A loss, various PVC methods have been developed (for a review, see Erlandsson et al., 2012). Importantly, different PVC algorithms may lead to different conclusions, with IY providing a theoretically more accurate PVC method compared to MG (Lu et al., 2021). This review encompasses clinical studies using various PVC methods across neurodegenerative disorders. Methodological differences persist even among studies that concentrate on the same pathology. For instance, Matuskey et al. (2020), Delva, Van Laere, et al. (2022), and Andersen et al. (2023) investigated synaptic density in Lewy body diseases, applying MG, RBV, and GTM PVC approaches respectively. In addition, other potential sources of bias include inconsistencies in PET imaging modalities, radiotracer selection, and modelling approaches. Furthermore, while some studies excluded participants using medications with known affinity for SV2A (e.g., Donepezil or Levetiracetam, Holland et al., 2020; Malpetti et al., 2023; Matuskey et al., 2020; Vanderlinden et al., 2022; Venkataraman et al., 2022), in the remaining studies, either medication use was not reported in general, or methods to account for it were not implemented. Since it is unknown how other centrally acting therapeutics may alter SV2A and synaptic density, this could potentially influence study results. For instance, long-term Levodopa use appears to affect

synaptic plasticity and may increase synaptic density through levodopa-induced dyskinesia (Calabresi et al., 2015; Hurley et al., 2005; Martin et al., 2023).

## **5. Summary, perspectives, and future directions**

This systematic review provides an extensive synthesis of the current state of *in vivo* SV2A PET imaging research of neurodegenerative disorders. We have discussed the main findings concerning group differences and clinical-cognitive correlations and explored relations between SV2A PET and other markers of pathology.

In summary, PET imaging of SV2A directly measures synaptic vesicles and serves as a proxy for synaptic density, thereby contributing to our understanding of the pathological processes preceding atrophy in neurodegeneration. Qualified SV2A radiotracers are available and validation studies have successfully identified their optimal compartment models and parametric approaches. Synaptic density appears to remain relatively preserved in the healthy ageing population, which may imply that synaptic density loss observed in neurodegenerative disorders primarily stem from the progressive disease itself, rather than from the natural ageing process. While the reduction of SV2A expression may not be specific to neurodegeneration in general, abnormal synaptic changes were identified in a large majority of neurodegenerative disorders. The consensus regarding the direction of disease-related changes in synaptic density, despite methodological variation among studies, suggests a strong association across different neurodegenerative disorders. The regional and temporal patterns of these changes could potentially offer valuable information that may aid in distinguishing between neurodegenerative conditions. Numerous studies underscore strong group differences in which anatomical patterns of synaptic density loss are found to be specific to brain regions linked to respective diseases. Longitudinally, studies reveal variations in the pace of synaptic density decline, and signal change seems to correlate with disease progression overall. Furthermore, the multitude of correlations found between synaptic density and clinical-cognitive functioning signifies its role in cognitive impairment, both globally and within specific domains. The complex relationship between synaptic density loss and other molecular pathologies may explain changes in cognition not attributable to GM atrophy.

### *5.1. Clinical relevance and the road ahead of in vivo synaptic density imaging*

Researchers are actively developing synapse-targeted therapies for neurodegenerative disorders (Peng et al., 2022). Preclinical studies suggest potential medications for AD may enhance synaptic density (Kaufman et al., 2015; Smith et al., 2018; Toyonaga et al., 2019). A promising avenue for future exploration is the use of SV2A imaging to monitor therapeutic responses and assess drug efficacy in human clinical trials. Initial progress in this direction has already been made (ID NCT 03493282).

Furthermore, delineating synaptic density pathology onto other imaging markers will provide more profound insights into *how* neurodegenerative diseases progress temporally, as well as elucidate the relative roles and causal relationships of different pathomechanisms (Drzezga et al., 2014). For instance, future research could involve employing structural equation modelling to gain insights into the temporal relationships between Tau, synaptic density, and cognitive impairment. Furthermore, investigating the link between synaptic density decline and A $\beta$ -toxicity could contribute to a more nuanced stratification of patient populations.

Studies with larger patient cohorts, implementing longitudinal approaches starting in preclinical phases, and direct comparisons of clinically resemblant diseases are desired. In pursuit of these objectives, standardising *in vivo* SV2A PET imaging protocols is considered ideal to tackle the ongoing methodological variability in the field and facilitating between-study comparisons. In light of this, it is relevant to further validate newly developed radiotracers, such as the readily available <sup>18</sup>F-labeled version of UCB-J, which are more openly available for clinical use. Additionally, validating PVC methods to control for cerebral atrophy is crucial for ensuring accurate and reliable SV2A PET imaging results (Becker et al., 2020; Mercier et al., 2017).

## 6. Concluding remarks

As a conclusion, SV2A radiotracers have enabled the characterisation of synaptic density loss in the living brain. SV2A PET imaging shows tremendous capability to provide novel insights into the aetiology of neurodegenerative disorders and great promise as a biomarker for synaptic density reduction. Of particular interest herein is its potential contribution to early and differential diagnosis. Furthermore, when combined with other imaging modalities and applied across the spectrum from normal ageing to dementia, SV2A PET imaging may promote efforts to develop disease-modifying therapies that target the preservation and restoration of synapses, making its range of clinical applications immense.

## 7. Competing interests

Unrelated to this work, JOB has received honoraria for work as DSMB chair or member for TauRx, Axon, Eisai, has acted as a consultant for Roche, and has received research support from Alliance Medical and Merck.

## 8. Funding

JOB receives infrastructural support from the Cambridge NIHR BRC. EM is supported by an Alzheimer's Society Junior Research Fellowship (443 AS JF 18017) and Lewy Body Society.

## 9. Declaration of Generative AI and AI-assisted technologies in the writing process

During the preparation of this work the author(s) used generative AI in order to improve the language and readability of the paper. After using this tool/service, the author(s) reviewed and edited the content as needed and take(s) full responsibility for the content of the publication.

## 10. Bibliography

- Adams, N. E., Jafarian, A., Perry, A., Rouse, M. A., Shaw, A. D., Murley, A. G., Cope, T. E., Bevan-Jones, W. R., Passamonti, L., Street, D., Holland, N., Nesbitt, D., Hughes, L. E., Friston, K. J., & Rowe, J. B. (2023). Neurophysiological consequences of synapse loss in progressive supranuclear palsy. *Brain: a Journal of Neurology*, *146*(6), 2584-2594. <https://doi.org/10.1093/brain/awac471>
- Andersen, K. B., Hansen, A. K., Damholdt, M. F., Horsager, J., Skjaerbaek, C., Gottrup, H., Klit, H., Schacht, A. C., Danielsen, E. H., Brooks, D. J., & Borghammer, P. (2021). Reduced Synaptic Density in Patients with Lewy Body Dementia: An [(11)C]UCB-J PET Imaging Study. *Movement Disorders*, *36*(9), 2057-2065. <https://doi.org/10.1002/mds.28617>
- Andersen, K. B., Hansen, A. K., Knudsen, K., Schacht, A. C., Damholdt, M. F., Brooks, D. J., & Borghammer, P. (2022). Healthy brain aging assessed with [(18)F]FDG and [(11)C]UCB-J PET. *Nuclear Medicine and Biology*, *112-113*(9), 52-58. <https://doi.org/10.1016/j.nucmedbio.2022.06.007>
- Andersen, K. B., Hansen, A. K., Schacht, A. C., Horsager, J., Gottrup, H., Klit, H., Danielsen, E. H., Poston, K. L., Pavese, N., Brooks, D. J., & Borghammer, P. (2023). Synaptic Density and Glucose Consumption in Patients with Lewy Body Diseases: An [(11)C]UCB-J and [(18)F]FDG PET Study. *Movement Disorders*, *38*(5), 796-805. <https://doi.org/10.1002/mds.29375>
- Bastin, C., Bahri, M. A., Meyer, F., Manard, M., Delhay, E., Plenevaux, A., Becker, G., Seret, A., Mella, C., Giacomelli, F., Degueldre, C., Balteau, E., Luxen, A., & Salmon, E. (2020). In vivo imaging of synaptic loss in Alzheimer's disease with [18F]UCB-H positron emission tomography. *European Journal of Nuclear Medicine and Molecular Imaging*, *47*(2), 390-402. <https://doi.org/10.1007/s00259-019-04461-x>
- Becker, G., Dammicco, S., Bahri, M. A., & Salmon, E. (2020). The Rise of Synaptic Density PET Imaging. *Molecules*, *25*(10). <https://doi.org/10.3390/molecules25102303>

- Cai, Z., Li, S., Matuskey, D., Nabulsi, N., & Huang, Y. (2019). PET imaging of synaptic density: A new tool for investigation of neuropsychiatric diseases. *Neuroscience Letters*, *691*, 44-50. <https://doi.org/10.1016/j.neulet.2018.07.038>
- Calabresi, P., Ghiglieri, V., Mazzocchi, P., Corbelli, I., & Picconi, B. (2015). Levodopa-induced plasticity: a double-edged sword in Parkinson's disease? *Philosophical Transactions of the Royal Society B*, *370*(1672). <https://doi.org/10.1098/rstb.2014.0184>
- Calhoun, M. E., Jucker, M., Martin, L. J., Thinakaran, G., Price, D. L., & Mouton, P. R. (1996). Comparative evaluation of synaptophysin-based methods for quantification of synapses. *Journal of Neurocytology*, *25*(12), 821-828. <https://doi.org/10.1007/BF02284844>
- Carapelle, E., Mundi, C., Cassano, T., & Avolio, C. (2020). Interaction between Cognitive Reserve and Biomarkers in Alzheimer Disease. *International Journal of Molecular Sciences*, *21*(17), 6279. <https://doi.org/10.3390/ijms21176279>
- Carson, R. E., Naganawa, M., Toyonaga, T., Koohsari, S., Yang, Y., Chen, M. K., Matuskey, D., & Finnema, S. J. (2022). Imaging of Synaptic Density in Neurodegenerative Disorders. *Journal of Nuclear Medicine*, *63*(Suppl 1), 60S-67S. <https://doi.org/10.2967/jnumed.121.263201>
- Chen, M. K., Mecca, A. P., Naganawa, M., Finnema, S. J., Toyonaga, T., Lin, S. F., Najafzadeh, S., Ropchan, J., Lu, Y., McDonald, J. W., Michalak, H. R., Nabulsi, N. B., Arnsten, A. F. T., Huang, Y., Carson, R. E., & van Dyck, C. H. (2018). Assessing Synaptic Density in Alzheimer Disease With Synaptic Vesicle Glycoprotein 2A Positron Emission Tomographic Imaging. *JAMA Neurology*, *75*(10), 1215-1224. <https://doi.org/10.1001/jamaneurol.2018.1836>
- Chen, M. K., Mecca, A. P., Naganawa, M., Gallezot, J. D., Toyonaga, T., Mondal, J., Finnema, S. J., Lin, S. F., O'Dell, R. S., McDonald, J. W., Michalak, H. R., Vander Wyk, B., Nabulsi, N. B., Huang, Y., Arnsten, A. F., van Dyck, C. H., & Carson, R. E. (2021). Comparison of [(11)C]UCB-J and [(18)F]FDG PET in Alzheimer's disease: A tracer kinetic modeling study. *Journal of Cerebral Blood Flow & Metabolism*, *41*(9), 2395-2409. <https://doi.org/10.1177/0271678X211004312>
- Chen, Z., Liao, G., Wan, N., He, Z., Chen, D., Tang, Z., Long, Z., Zou, G., Peng, L., Wan, L., Wang, C., Peng, H., Shi, Y., Tang, Y., Li, J., Li, Y., Long, T., Hou, X., He, L., . . . Jiang, H. (2023). Synaptic Loss in Spinocerebellar Ataxia Type 3 Revealed by SV2A Positron Emission Tomography. *Movement Disorders*, *38*(6), 978-989. <https://doi.org/10.1002/mds.29395>
- Chetelat, G., Landeau, B., Salmon, E., Yakushev, I., Bahri, M. A., Mezenge, F., Perrotin, A., Bastin, C., Manrique, A., Scheurich, A., Scheckenberger, M., Desgranges, B., Eustache, F., & Fellgiebel, A. (2013). Relationships between brain metabolism decrease in normal aging and changes in structural and functional connectivity. *NeuroImage*, *76*, 167-177. <https://doi.org/10.1016/j.neuroimage.2013.03.009>
- Coomans, E. M., Schoonhoven, D. N., Tuncel, H., Verfaillie, S. C. J., Wolters, E. E., Boellaard, R., Ossenkoppele, R., den Braber, A., Scheper, W., Schober, P., Sweeney, S. P., Ryan, J. M., Schuit, R. C., Windhorst, A. D., Barkhof, F., Scheltens, P., Golla, S. S. V., Hillebrand, A., Gouw, A. A., & van Berckel, B. N. M. (2021). In vivo tau pathology is associated with synaptic loss and altered synaptic function. *Alzheimer's Research & Therapy*, *13*(1), 35. <https://doi.org/10.1186/s13195-021-00772-0>
- Delva, A., Michiels, L., Koole, M., Van Laere, K., & Vandenberghe, W. (2022). Synaptic Damage and Its Clinical Correlates in People With Early Huntington Disease: A PET Study. *Neurology*, *98*(1), e83-e94. <https://doi.org/10.1212/WNL.0000000000012969>
- Delva, A., Van Laere, K., & Vandenberghe, W. (2022). Longitudinal Positron Emission Tomography Imaging of Presynaptic Terminals in Early Parkinson's Disease. *Movement Disorders*, *37*(9), 1883-1892. <https://doi.org/10.1002/mds.29148>
- Delva, A., Van Weehaeghe, D., Koole, M., Van Laere, K., & Vandenberghe, W. (2020). Loss of Presynaptic Terminal Integrity in the Substantia Nigra in Early Parkinson's Disease. *Movement Disorders*, *35*(11), 1977-1986. <https://doi.org/10.1002/mds.28216>
- Dierckx, R. A. J. O., Otte, A., de Vries, E. F. J., van Waarde, A., & Leenders, K. L. (2021). *PET and SPECT in Neurology* (2 ed.). Springer Nature Switzerland.
- Drzezga, A., Barthel, H., Minoshima, S., & Sabri, O. (2014). Potential Clinical Applications of PET/MR Imaging in Neurodegenerative Diseases. *Journal of Nuclear Medicine*, *55*(Supplement 2), 47S-55S. <https://doi.org/10.2967/jnumed.113.129254>

- Dugger, B. N., & Dickson, D. W. (2017). Pathology of Neurodegenerative Diseases. *Cold Spring Harbor Perspectives in Biology*, 9(7). <https://doi.org/10.1101/cshperspect.a028035>
- Erlandsson, K., Buvat, I., Pretorius, P. H., Thomas, B. A., & Hutton, B. F. (2012). A review of partial volume correction techniques for emission tomography and their applications in neurology, cardiology and oncology. *Physics in Medicine & Biology*, 57(21), R119-159. <https://doi.org/10.1088/0031-9155/57/21/R119>
- Fang, X. T., Toyonaga, T., Hillmer, A. T., Matuskey, D., Holmes, S. E., Radhakrishnan, R., Mecca, A. P., van Dyck, C. H., D'Souza, D. C., Esterlis, I., Worhunsky, P. D., & Carson, R. E. (2021). Identifying brain networks in synaptic density PET ((11)C-UCB-J) with independent component analysis. *NeuroImage*, 237, 118167. <https://doi.org/10.1016/j.neuroimage.2021.118167>
- Fang, X. T., Volpi, T., Holmes, S. E., Esterlis, I., Carson, R. E., & Worhunsky, P. D. (2023). Linking resting-state network fluctuations with systems of coherent synaptic density: A multimodal fMRI and (11)C-UCB-J PET study. *Frontiers in Human Neuroscience*, 17, 1124254. <https://doi.org/10.3389/fnhum.2023.1124254>
- Feng, G., Xiao, F., Lu, Y., Huang, Z., Yuan, J., Xiao, Z., Xi, Z., & Wang, X. (2009). Down-regulation synaptic vesicle protein 2A in the anterior temporal neocortex of patients with intractable epilepsy. *Journal of Molecular Neuroscience*, 39(3), 354-359. <https://doi.org/10.1007/s12031-009-9288-2>
- Finnema, S. J., Nabulsi, N. B., Eid, T., Detyniecki, K., Lin, S. F., Chen, M. K., Dhaher, R., Matuskey, D., Baum, E., Holden, D., Spencer, D. D., Mercier, J., Hannestad, J., Huang, Y., & Carson, R. E. (2016). Imaging synaptic density in the living human brain. *Science Translational Medicine*, 8(348), 348ra396. <https://doi.org/10.1126/scitranslmed.aaf6667>
- Finnema, S. J., Nabulsi, N. B., Mercier, J., Lin, S. F., Chen, M. K., Matuskey, D., Gallezot, J. D., Henry, S., Hannestad, J., Huang, Y., & Carson, R. E. (2018). Kinetic evaluation and test-retest reproducibility of [(11)C]UCB-J, a novel radioligand for positron emission tomography imaging of synaptic vesicle glycoprotein 2A in humans. *Journal of Cerebral Blood Flow & Metabolism*, 38(11), 2041-2052. <https://doi.org/10.1177/0271678X17724947>
- Finnema, S. J., Toyonaga, T., Detyniecki, K., Chen, M. K., Dias, M., Wang, Q., Lin, S. F., Naganawa, M., Gallezot, J. D., Lu, Y., Nabulsi, N. B., Huang, Y., Spencer, D. D., & Carson, R. E. (2020). Reduced synaptic vesicle protein 2A binding in temporal lobe epilepsy: A [(11)C]UCB-J positron emission tomography study. *Epilepsia*, 61(10), 2183-2193. <https://doi.org/10.1111/epi.16653>
- Fujimoto, T., Matsumoto, T., Fujita, S., Takeuchi, K., Nakamura, K., Mitsuyama, Y., & Kato, N. (2008). Changes in glucose metabolism due to aging and gender-related differences in the healthy human brain. *Psychiatry Research*, 164(1), 58-72. <https://doi.org/10.1016/j.psychresns.2006.12.014>
- Gillard, M., Fuks, B., Leclercq, K., & Matagne, A. (2011). Binding characteristics of brivaracetam, a selective, high affinity SV2A ligand in rat, mouse and human brain: relationship to anti-convulsant properties. *European Journal of Pharmacology*, 664(1-3), 36-44. <https://doi.org/10.1016/j.ejphar.2011.04.064>
- Heeman, F., Yaqub, M., Hendriks, J., van Berckel, B. N. M., Collij, L. E., Gray, K. R., Manber, R., Wolz, R., Garibotto, V., Wimberley, C., Ritchie, C., Barkhof, F., Gispert, J. D., Vallez Garcia, D., Lopes Alves, I., Lammertsma, A. A., & Consortium, A. (2022). Impact of cerebral blood flow and amyloid load on SUVR bias. *EJNMMI Res*, 12(1), 29. <https://doi.org/10.1186/s13550-022-00898-8>
- Herholz, K., Carter, S. F., & Jones, M. (2007). Positron emission tomography imaging in dementia. *The British Journal of Radiology*, 80 Spec No 2(SPEC. ISS. 2), S160-167. <https://doi.org/10.1259/bjr/97295129>
- Holland, N., Jones, P. S., Savulich, G., Naessens, M., Malpetti, M., Whiteside, D. J., Street, D., Swann, P., Hong, Y. T., Fryer, T. D., Rittman, T., Mulroy, E., Aigbirhio, F. I., Bhatia, K. P., O'Brien, J. T., & Rowe, J. B. (2023). Longitudinal Synaptic Loss in Primary Tauopathies: An In Vivo [(11)C]UCB-J Positron Emission Tomography Study. *Movement Disorders*, 38(7), 1316-1326. <https://doi.org/10.1002/mds.29421>

- Holland, N., Jones, P. S., Savulich, G., Wiggins, J. K., Hong, Y. T., Fryer, T. D., Manavaki, R., Sephton, S. M., Boros, I., Malpetti, M., Hezemans, F. H., Aigbirhio, F. I., Coles, J. P., O'Brien, J., & Rowe, J. B. (2020). Synaptic Loss in Primary Tauopathies Revealed by [(11)C]UCB-J Positron Emission Tomography. *Movement Disorders*, 35(10), 1834-1842. <https://doi.org/10.1002/mds.28188>
- Holland, N., Malpetti, M., Rittman, T., Mak, E. E., Passamonti, L., Kaalund, S. S., Hezemans, F. H., Jones, P. S., Savulich, G., Hong, Y. T., Fryer, T. D., Aigbirhio, F. I., O'Brien, J. T., & Rowe, J. B. (2022). Molecular pathology and synaptic loss in primary tauopathies: an 18F-AV-1451 and 11C-UCB-J PET study. *Brain: a Journal of Neurology*, 145(1), 340-348. <https://doi.org/10.1093/brain/awab282>
- Hsieh, T. C., Lin, W. Y., Ding, H. J., Sun, S. S., Wu, Y. C., Yen, K. Y., & Kao, C. H. (2012). Sex- and age-related differences in brain FDG metabolism of healthy adults: an SPM analysis. *Journal of Neuroimaging*, 22(1), 21-27. <https://doi.org/10.1111/j.1552-6569.2010.00543.x>
- Hurley, M. J., Jackson, M. J., Smith, L. A., Rose, S., & Jenner, P. (2005). Immunoautoradiographic analysis of NMDA receptor subunits and associated postsynaptic density proteins in the brain of dyskinetic MPTP-treated common marmosets. *European Journal of Neuroscience*, 21(12), 3240-3250. <https://doi.org/10.1111/j.1460-9568.2005.04169.x>
- Iaccarino, L., Sala, A., Caminiti, S. P., & Perani, D. (2017). The emerging role of PET imaging in dementia. *F1000Research*, 6, 1830. <https://doi.org/10.12688/f1000research.11603.1>
- Jack, C. R., Jr., Bennett, D. A., Blennow, K., Carrillo, M. C., Dunn, B., Haeberlein, S. B., Holtzman, D. M., Jagust, W., Jessen, F., Karlawish, J., Liu, E., Molinuevo, J. L., Montine, T., Phelps, C., Rankin, K. P., Rowe, C. C., Scheltens, P., Siemers, E., Snyder, H. M., . . . Contributors. (2018). NIA-AA Research Framework: Toward a biological definition of Alzheimer's disease. *Alzheimer's & Dementia*, 14(4), 535-562. <https://doi.org/10.1016/j.jalz.2018.02.018>
- Kaufman, A. C., Salazar, S. V., Haas, L. T., Yang, J., Kostylev, M. A., Jeng, A. T., Robinson, S. A., Gunther, E. C., van Dyck, C. H., Nygaard, H. B., & Strittmatter, S. M. (2015). Fyn inhibition rescues established memory and synapse loss in Alzheimer mice. *Annals of Neurology*, 77(6), 953-971. <https://doi.org/10.1002/ana.24394>
- Koole, M., van Aalst, J., Devrome, M., Mertens, N., Serdons, K., Lacroix, B., Mercier, J., Sciberras, D., Maguire, P., & Van Laere, K. (2019). Quantifying SV2A density and drug occupancy in the human brain using [(11)C]UCB-J PET imaging and subcortical white matter as reference tissue. *European Journal of Nuclear Medicine and Molecular Imaging*, 46(2), 396-406. <https://doi.org/10.1007/s00259-018-4119-8>
- Lepeta, K., Lourenco, M. V., Schweitzer, B. C., Martino Adami, P. V., Banerjee, P., Catuara-Solarz, S., de La Fuente Revenga, M., Guillem, A. M., Haidar, M., Ijomone, O. M., Nadorp, B., Qi, L., Perera, N. D., Refsgaard, L. K., Reid, K. M., Sabbar, M., Sahoo, A., Schaefer, N., Sheean, R. K., . . . Seidenbecher, C. (2016). Synaptopathies: synaptic dysfunction in neurological disorders - A review from students to students. *Journal of Neurochemistry*, 138(6), 785-805. <https://doi.org/10.1111/jnc.13713>
- Li, S., Naganawa, M., Pracitto, R., Najafzadeh, S., Holden, D., Henry, S., Matuskey, D., Emery, P. R., Cai, Z., Ropchan, J., Nabulsi, N., Carson, R. E., & Huang, Y. (2021). Assessment of test-retest reproducibility of [(18)F]SynVesT-1, a novel radiotracer for PET imaging of synaptic vesicle glycoprotein 2A. *European Journal of Nuclear Medicine and Molecular Imaging*, 48(5), 1327-1338. <https://doi.org/10.1007/s00259-020-05149-3>
- Lu, Y., Toyonaga, T., Naganawa, M., Gallezot, J. D., Chen, M. K., Mecca, A. P., van Dyck, C. H., & Carson, R. E. (2021). Partial volume correction analysis for (11)C-UCB-J PET studies of Alzheimer's disease. *NeuroImage*, 238(April), 118248. <https://doi.org/10.1016/j.neuroimage.2021.118248>
- Luo, J., Norris, R. H., Gordon, S. L., & Nithianantharajah, J. (2018). Neurodevelopmental synaptopathies: Insights from behaviour in rodent models of synapse gene mutations. *Progress in Neuro-Psychopharmacology and Biological Psychiatry*, 84(Pt B), 424-439. <https://doi.org/10.1016/j.pnpbp.2017.12.001>
- Lynch, B. A., Lambeng, N., Nocka, K., Kensel-Hammes, P., Bajjalieh, S. M., Matagne, A., & Fuks, B. (2004). The synaptic vesicle protein SV2A is the binding site for the antiepileptic drug

- levetiracetam [Article]. *Proceedings of the National Academy of Sciences*, 101(26), 9861-9866. <https://doi.org/10.1073/pnas.0308208101>
- Mak, E., Holland, N., Jones, P. S., Savulich, G., Low, A., Malpetti, M., Kaalund, S. S., Passamonti, L., Rittman, T., Romero-Garcia, R., Manavaki, R., Williams, G. B., Hong, Y. T., Fryer, T. D., Aigbirhio, F. I., O'Brien, J. T., & Rowe, J. B. (2021). In vivo coupling of dendritic complexity with presynaptic density in primary tauopathies. *Neurobiology of Aging*, 101, 187-198. <https://doi.org/10.1016/j.neurobiolaging.2021.01.016>
- Malpetti, M., Holland, N., Jones, P. S., Ye, R., Cope, T. E., Fryer, T. D., Hong, Y. T., Savulich, G., Rittman, T., Passamonti, L., Mak, E., Aigbirhio, F. I., O'Brien, J. T., & Rowe, J. B. (2021). Synaptic density in carriers of C9orf72 mutations: a [(11)C]UCB-J PET study. *Annals of Clinical and Translational Neurology*, 8(7), 1515-1523. <https://doi.org/10.1002/acn3.51407>
- Malpetti, M., Jones, P. S., Cope, T. E., Holland, N., Naessens, M., Rouse, M. A., Rittman, T., Savulich, G., Whiteside, D. J., Street, D., Fryer, T. D., Hong, Y. T., Milicevic Sephton, S., Aigbirhio, F. I., JT, O. B., & Rowe, J. B. (2023). Synaptic Loss in Frontotemporal Dementia Revealed by [(11)C]UCB-J Positron Emission Tomography. *Annals of Neurology*, 93(1), 142-154. <https://doi.org/10.1002/ana.26543>
- Malpetti, M., Simon Jones, P., Cope, T. E., Holland, N., Naessens, M., Rouse, M. A., Savulich, G., Fryer, T. D., Hong, Y. T., Sephton, S. M., Aigbirhio, F. I., O'Brien, J. T., & Rowe, J. B. (2022). Synaptic loss in behavioural variant frontotemporal dementia revealed by [11C]UCB-J PET. *medRxiv*. <https://doi.org/10.1101/2022.01.30.22270123>
- Mansur, A., Rabiner, E. A., Comley, R. A., Lewis, Y., Middleton, L. T., Huiban, M., Passchier, J., Tsukada, H., & Gunn, R. N. (2020). Characterization of 3 PET Tracers for Quantification of Mitochondrial and Synaptic Function in Healthy Human Brain: (18)F-BCPP-EF, (11)C-SA-4503, and (11)C-UCB-J. *The Journal of Nuclear Medicine*, 61(1), 96-103. <https://doi.org/doi:10.2967/jnumed.119.228080>
- Martin, S. L., Uribe, C., & Strafella, A. P. (2023). PET imaging of synaptic density in Parkinsonian disorders. *Journal of Neuroscience Research*. <https://doi.org/10.1002/jnr.25253>
- Matuskey, D., Tinaz, S., Wilcox, K. C., Naganawa, M., Toyonaga, T., Dias, M., Henry, S., Pittman, B., Ropchan, J., Nabulsi, N., Suridjan, I., Comley, R. A., Huang, Y., Finnema, S. J., & Carson, R. E. (2020). Synaptic Changes in Parkinson Disease Assessed with in vivo Imaging. *Annals of Neurology*, 87(3), 329-338. <https://doi.org/10.1002/ana.25682>
- Mecca, A. P., Chen, M. K., O'Dell, R. S., Naganawa, M., Toyonaga, T., Godek, T. A., Harris, J. E., Bartlett, H. H., Zhao, W., Banks, E. R., Ni, G. S., Rogers, K., Gallezot, J. D., Ropchan, J., Emery, P. R., Nabulsi, N. B., Vander Wyk, B. C., Arnsten, A. F. T., Huang, Y., . . . van Dyck, C. H. (2022). Association of entorhinal cortical tau deposition and hippocampal synaptic density in older individuals with normal cognition and early Alzheimer's disease. *Neurobiology of Aging*, 111, 44-53. <https://doi.org/10.1016/j.neurobiolaging.2021.11.004>
- Mecca, A. P., Chen, M. K., O'Dell, R. S., Naganawa, M., Toyonaga, T., Godek, T. A., Harris, J. E., Bartlett, H. H., Zhao, W., Nabulsi, N. B., Wyk, B. C. V., Varma, P., Arnsten, A. F. T., Huang, Y., Carson, R. E., & van Dyck, C. H. (2020). In vivo measurement of widespread synaptic loss in Alzheimer's disease with SV2A PET. *Alzheimer's & Dementia*, 16(7), 974-982. <https://doi.org/10.1002/alz.12097>
- Mecca, A. P., O'Dell, R. S., Sharp, E. S., Banks, E. R., Bartlett, H. H., Zhao, W., Lipior, S., Diepenbrock, N. G., Chen, M. K., Naganawa, M., Toyonaga, T., Nabulsi, N. B., Vander Wyk, B. C., Arnsten, A. F. T., Huang, Y., Carson, R. E., & van Dyck, C. H. (2022). Synaptic density and cognitive performance in Alzheimer's disease: A PET imaging study with [(11)C]UCB-J. *Alzheimer's & Dementia*, 18(12), 2527-2536. <https://doi.org/10.1002/alz.12582>
- Mercier, J., Archen, L., Bollu, V., Carre, S., Evrard, Y., Jnoff, E., Kenda, B., Lallemand, B., Michel, P., Montel, F., Moureau, F., Price, N., Quesnel, Y., Sauvage, X., Valade, A., & Provins, L. (2014). Discovery of heterocyclic nonacetamide synaptic vesicle protein 2A (SV2A) ligands with single-digit nanomolar potency: opening avenues towards the first SV2A positron emission tomography (PET) ligands. *ChemMedChem*, 9(4), 693-698. <https://doi.org/10.1002/cmde.201300482>

- Mercier, J., Provins, L., & Valade, A. (2017). Discovery and development of SV2A PET tracers: Potential for imaging synaptic density and clinical applications. *Drug Discovery Today: Technologies*, 25, 45-52. <https://doi.org/10.1016/j.ddtec.2017.11.003>
- Mertens, N., Maguire, R. P., Serdons, K., Lacroix, B., Mercier, J., Sciberras, D., Van Laere, K., & Koole, M. (2020). Validation of Parametric Methods for [(11)C]UCB-J PET Imaging Using Subcortical White Matter as Reference Tissue. *Molecular Imaging and Biology*, 22(2), 444-452. <https://doi.org/10.1007/s11307-019-01387-6>
- Michiels, L., Delva, A., van Aalst, J., Ceccarini, J., Vandenberghe, W., Vandenbulcke, M., Koole, M., Lemmens, R., & Laere, K. V. (2021). Synaptic density in healthy human aging is not influenced by age or sex: a (11)C-UCB-J PET study. *NeuroImage*, 232, 117877. <https://doi.org/10.1016/j.neuroimage.2021.117877>
- Naganawa, M., Li, S., Nabulsi, N., Henry, S., Zheng, M. Q., Pracitto, R., Cai, Z., Gao, H., Kapinos, M., Labaree, D., Matuskey, D., Huang, Y., & Carson, R. E. (2021). First-in-Human Evaluation of (18)F-SynVesT-1, a Radioligand for PET Imaging of Synaptic Vesicle Glycoprotein 2A. *J Nucl Med*, 62(4), 561-567. <https://doi.org/10.2967/jnumed.120.249144>
- O'Dell, R., Mecca, A., Chen, M. K., Naganawa, M., Toyonaga, T., Lu, Y., Godek, T., Harris, J., Bartlett, H., Banks, E., Kominek, V., Zhao, W., Nabulsi, N., Ropchan, J., Ye, Y., Vander Wyk, B., Huang, Y., Arnsten, A., Carson, R., & van Dyck, C. (2021). Association of Aβ deposition and regional synaptic density in early Alzheimer's disease: a PET imaging study with [11C]UCB-J. *Am. J. Geriatr. Psychiatry*, 29(4), S37-S40. <https://doi.org/10.1016/j.jagp.2021.01.034>
- Page, M. J., Moher, D., Bossuyt, P. M., Boutron, I., Hoffmann, T. C., Mulrow, C. D., Shamseer, L., Tetzlaff, J. M., Akl, E. A., Brennan, S. E., Chou, R., Glanville, J., Grimshaw, J. M., Hrobjartsson, A., Lalu, M. M., Li, T., Loder, E. W., Mayo-Wilson, E., McDonald, S., . . . McKenzie, J. E. (2021). PRISMA 2020 explanation and elaboration: updated guidance and exemplars for reporting systematic reviews. *British Medical Journal*, 372, n160. <https://doi.org/10.1136/bmj.n160>
- Peng, L., Bestard-Lorigados, I., & Song, W. (2022). The synapse as a treatment avenue for Alzheimer's Disease. *Molecular Psychiatry*, 27(7), 2940-2949. <https://doi.org/10.1038/s41380-022-01565-z>
- Rabiner, E. A. (2018). Imaging Synaptic Density: A Different Look at Neurologic Diseases. *J Nucl Med*, 59(3), 380-381. <https://doi.org/10.2967/jnumed.117.198317>
- Ritchie, K., Carriere, I., Su, L., O'Brien, J. T., Lovestone, S., Wells, K., & Ritchie, C. W. (2017). The midlife cognitive profiles of adults at high risk of late-onset Alzheimer's disease: The PREVENT study. *Alzheimer's & Dementia*, 13(10), 1089-1097. <https://doi.org/10.1016/j.jalz.2017.02.008>
- Rizzoli, S. O., & Betz, W. J. (2005). Synaptic vesicle pools. *Nature Reviews Neuroscience*, 6(1), 57-69. <https://doi.org/10.1038/nrn1583>
- Rossano, S., Toyonaga, T., Finnema, S. J., Naganawa, M., Lu, Y., Nabulsi, N., Ropchan, J., De Bruyn, S., Otoul, C., Stockis, A., Nicolas, J. M., Martin, P., Mercier, J., Huang, Y., Maguire, R. P., & Carson, R. E. (2020). Assessment of a white matter reference region for (11)C-UCB-J PET quantification. *Journal of Cerebral Blood Flow & Metabolism*, 40(9), 1890-1901. <https://doi.org/10.1177/0271678X19879230>
- Rub, U., Seidel, K., Heinsen, H., Vonsattel, J. P., den Dunnen, W. F., & Korf, H. W. (2016). Huntington's disease (HD): the neuropathology of a multisystem neurodegenerative disorder of the human brain. *Brain Pathology*, 26(6), 726-740. <https://doi.org/10.1111/bpa.12426>
- Salmon, E., Bahri, M. A., Plenevaux, A., Becker, G., Seret, A., Delhay, E., Degueldre, C., Balteau, E., Lemaire, C., Luxen, A., & Bastin, C. (2021). In vivo exploration of synaptic projections in frontotemporal dementia. *Scientific Reports*, 11(1), 16092. <https://doi.org/10.1038/s41598-021-95499-1>
- Serrano, M. E., Kim, E., Petrinovic, M. M., Turkheimer, F., & Cash, D. (2022). Imaging Synaptic Density: The Next Holy Grail of Neuroscience? *Frontiers in Neuroscience*, 16, 796129. <https://doi.org/10.3389/fnins.2022.796129>

- Smith, L. M., Zhu, R., & Strittmatter, S. M. (2018). Disease-modifying benefit of Fyn blockade persists after washout in mouse Alzheimer's model. *Neuropharmacology*, *130*, 54-61. <https://doi.org/10.1016/j.neuropharm.2017.11.042>
- Topcuoglu, E. S., Akdemir, U. O., & Atay, L. O. (2022a). What is New in Nuclear Medicine Imaging for Dementia. *Nöropsikiyatri arşivi*, *59*(Suppl 1), S17-S23. <https://doi.org/10.29399/npa.28155>
- Topcuoglu, E. S., Akdemir, U. O., & Atay, L. O. (2022b). What is New in Nuclear Medicine Imaging for Dementia. *Archives of Neuropsychiatry*, *59*(Suppl 1), S17-S23. <https://doi.org/10.29399/npa.28155>
- Toyonaga, T., Smith, L. M., Finnema, S. J., Gallezot, J. D., Naganawa, M., Bini, J., Mulnix, T., Cai, Z., Ropchan, J., Huang, Y., Strittmatter, S. M., & Carson, R. E. (2019). In Vivo Synaptic Density Imaging with (11)C-UCB-J Detects Treatment Effects of Saracatinib in a Mouse Model of Alzheimer Disease. *J Nucl Med*, *60*(12), 1780-1786. <https://doi.org/10.2967/jnumed.118.223867>
- Tuncel, H., Boellaard, R., Coomans, E. M., de Vries, E. F., Glaudemans, A. W., Feltes, P. K., Garcia, D. V., Verfaillie, S. C., Wolters, E. E., Sweeney, S. P., Ryan, J. M., Ivarsson, M., Lynch, B. A., Schober, P., Scheltens, P., Schuit, R. C., Windhorst, A. D., De Deyn, P. P., van Berckel, B. N., & Golla, S. S. (2021). Kinetics and 28-day test-retest repeatability and reproducibility of [(11)C]UCB-J PET brain imaging. *Journal of Cerebral Blood Flow & Metabolism*, *41*(6), 1338-1350. <https://doi.org/10.1177/0271678X20964248>
- Tuncel, H., Boellaard, R., Coomans, E. M., Hollander-Meeuwse, M. D., de Vries, E. F. J., Glaudemans, A., Feltes, P. K., Garcia, D. V., Verfaillie, S. C. J., Wolters, E. E., Sweeney, S. P., Ryan, J. M., Ivarsson, M., Lynch, B. A., Schober, P., Scheltens, P., Schuit, R. C., Windhorst, A. D., De Deyn, P. P., . . . Golla, S. S. V. (2022). Validation and test-retest repeatability performance of parametric methods for [(11)C]UCB-J PET. *EJNMMI Res*, *12*(1), 3. <https://doi.org/10.1186/s13550-021-00874-8>
- van Aalst, J., Ceccarini, J., Sunaert, S., Dupont, P., Koole, M., & Van Laere, K. (2021). In vivo synaptic density relates to glucose metabolism at rest in healthy subjects, but is strongly modulated by regional differences. *Journal of Cerebral Blood Flow & Metabolism*, *41*(8), 1978-1987. <https://doi.org/10.1177/0271678X20981502>
- van Waarde, A., Marcolini, S., de Deyn, P. P., & Dierckx, R. (2021). PET Agents in Dementia: An Overview. *Seminars in Nuclear Medicine*, *51*(3), 196-229. <https://doi.org/10.1053/j.semnuclmed.2020.12.008>
- Vanderlinden, G., Ceccarini, J., Vande Castele, T., Michiels, L., Lemmens, R., Triau, E., Serdons, K., Tournoy, J., Koole, M., Vandenbulcke, M., & Van Laere, K. (2022). Spatial decrease of synaptic density in amnesic mild cognitive impairment follows the tau build-up pattern. *Molecular Psychiatry*, *27*(10), 4244-4251. <https://doi.org/10.1038/s41380-022-01672-x>
- Vanhaute, H., Ceccarini, J., Michiels, L., Koole, M., Sunaert, S., Lemmens, R., Triau, E., Emsell, L., Vandenbulcke, M., & Van Laere, K. (2020). In vivo synaptic density loss is related to tau deposition in amnesic mild cognitive impairment. *Neurology*, *95*(5), e545-e553. <https://doi.org/10.1212/WNL.0000000000009818>
- Venkataraman, A. V., Mansur, A., Rizzo, G., Bishop, C., Lewis, Y., Kocagoncu, E., Lingford-Hughes, A., Huiban, M., Passchier, J., Rowe, J. B., Tsukada, H., Brooks, D. J., Martarello, L., Comley, R. A., Chen, L., Schwarz, A. J., Hargreaves, R., Gunn, R. N., Rabiner, E. A., & Matthews, P. M. (2022). Widespread cell stress and mitochondrial dysfunction occur in patients with early Alzheimer's disease. *Science Translational Medicine*, *14*(658), eabk1051. <https://doi.org/10.1126/scitranslmed.abk1051>
- Vonsattel, J. P. G., Keller, C., & Cortes Ramirez, E. P. (2011). Chapter 4 - Huntington's disease – neuropathology. In W. J. Weiner & E. Tolosa (Eds.), *Handbook of Clinical Neurology* (Vol. 100, pp. 83-100). Elsevier. <https://doi.org/https://doi.org/10.1016/B978-0-444-52014-2.00004-5>
- Wang, R., Liu, H., Toyonaga, T., Shi, L., Wu, J., Onofrey, J. A., Tsai, Y. J., Naganawa, M., Ma, T., Liu, Y., Chen, M. K., Mecca, A. P., O'Dell, R. S., van Dyck, C. H., Carson, R. E., & Liu, C. (2021). Generation of synthetic PET images of synaptic density and amyloid from (18) F-

- FDG images using deep learning. *Medical physics*, 48(9), 5115-5129. <https://doi.org/10.1002/mp.15073>
- Wilson, H., Pagano, G., de Natale, E. R., Mansur, A., Caminiti, S. P., Polychronis, S., Middleton, L. T., Price, G., Schmidt, K. F., Gunn, R. N., Rabiner, E. A., & Politis, M. (2020). Mitochondrial Complex 1, Sigma 1, and Synaptic Vesicle 2A in Early Drug-Naive Parkinson's Disease. *Movement Disorders*, 35(8), 1416-1427. <https://doi.org/10.1002/mds.28064>
- Wu, M., Zhang, M., Yin, X., Chen, K., Hu, Z., Zhou, Q., Cao, X., Chen, Z., & Liu, D. (2021). The role of pathological tau in synaptic dysfunction in Alzheimer's diseases. *Translational Neurodegeneration*, 10(1), 45. <https://doi.org/10.1186/s40035-021-00270-1>
- Yoshizawa, H., Gazes, Y., Stern, Y., Miyata, Y., & Uchiyama, S. (2014). Characterizing the normative profile of 18F-FDG PET brain imaging: sex difference, aging effect, and cognitive reserve. *Psychiatry Research*, 221(1), 78-85. <https://doi.org/10.1016/j.psychres.2013.10.009>
- Zanderigo, F., Ogden, R. T., & Parsey, R. V. (2013). Reference region approaches in PET: a comparative study on multiple radioligands. *Journal of Cerebral Blood Flow & Metabolism*, 33(6), 888-897. <https://doi.org/10.1038/jcbfm.2013.26>
- Zhang, H., Schneider, T., Wheeler-Kingshott, C. A., & Alexander, D. C. (2012). NODDI: practical in vivo neurite orientation dispersion and density imaging of the human brain. *NeuroImage*, 61(4), 1000-1016. <https://doi.org/10.1016/j.neuroimage.2012.03.072>
- Zhang, J., Wang, J., Xu, X., You, Z., Huang, Q., Huang, Y., Guo, Q., Guan, Y., Zhao, J., Liu, J., Xu, W., Deng, Y., Xie, F., & Li, B. (2023). In vivo synaptic density loss correlates with impaired functional and related structural connectivity in Alzheimer's disease. *Journal of Cerebral Blood Flow & Metabolism*, 43(6), 977-988. <https://doi.org/10.1177/0271678X231153730>